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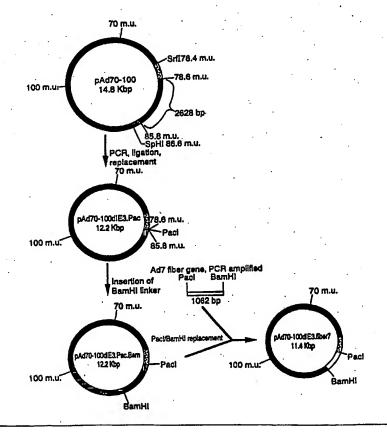
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(54) Title: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

#### (57) Abstract

The present invention provides a chimeric adenoviral coat protein (particularly a chimeric adenovirus hexon protein). The chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type adenovirus coat protein.



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WO 98/40509 PCT/US98/05033

# CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME

#### TECHNICAL FIELD OF THE INVENTION

The present invention relates to a chimeric adenoviral coat protein and a recombinant adenovirus comprising same. In particular, the invention provides a chimeric adenoviral hexon protein and a recombinant adenovirus comprising the chimeric adenoviral hexon protein. Such a recombinant adenovirus can be employed inter alia in gene therapy.

#### BACKGROUND OF THE INVENTION

In vivo gene therapy is a strategy in which nucleic acid, usually in the form of DNA, is administered to modify the genetic repertoire of target cells for . therapeutic purposes. This can be accomplished efficiently using a recombinant adenoviral vector encoding a so-called "therapeutic gene". A therapeutic gene is generally considered a gene that corrects or compensates for an underlying protein deficit or, alternately, a gene that is capable of down-regulating a particular gene, or counteracting the negative effects of its encoded product, in a given disease state or syndrome. Recombinant adenoviral vectors have been used to transfer one or more recombinant genes to diseased cells or tissues in need of treatment. As reviewed by Crystal, <u>Science</u>, <u>270</u>, 404-410 (1995), such vectors are preferred over other vectors commonly employed for gene therapy (e.g., retroviral vectors) since adenoviral vectors can be produced in high titers (i.e., up to 10<sup>13</sup> viral particles/ml), and they efficiently transfer genes to nonreplicating, as well as replicating, cells. Moreover, adenoviral vectors are additionally preferred based on their normal tropism for

the respiratory epithelium in cases where the targeted tissue for somatic gene therapy is the lung, as well as for other reasons (see, e.g., Straus, In Adenoviruses, Plenan Press, New York, NY, 451-496 (1984)); Horwitz et al., In Virology, 2nd Ed., Fields et al., eds., Raven Press, New York, NY, 1679-1721 (1990); Berkner, BioTechniques, 6, 616 (1988); Chanock et al., JAMA, 195, 151 (1966); Haj-Ahmad et al., J. Virol., 57, 267 (1986); and Ballay et al., EMBO, 4, 3861 (1985)).

There are 49 human adenoviral serotypes, categorized into 6 subgenera (A through F) based on nucleic acid comparisons, fiber protein characteristics, and biological properties (Crawford-Miksza et al., J. Virol., 70, 1836-1844 (1996)). The group C viruses (e.g., serotypes 2 and 5, or Ad2 and Ad5) are well characterized. It is these serotypes that currently are employed for gene transfer studies, including human gene therapy trials (see, e.g., Rosenfeld et al., Science, 252, 431-434 (1991); Rosenfeld et al., Cell, 68, 143-155 (1992); Zabner, Cell, 75, 207-216 (1993); Crystal et al., Nat. Gen., 8, 42-51 (1994); Yei et al., Gene Therapy, 1, 192-200 (1994); Chen et al., Proc. Natl. Acad. Sci., 91, 3054-3057 (1994); Yang et al., Nat. Gen., 7, 362-369 (1994); Zabner et al., Nat. Gen., 6, 75-83 (1994)). Other groups and serotypes include, but are not limited to: group A (e.g., serotypes 12 and 31), group B (e.g., serotypes 3 and 7), group D (e.g., serotypes 8 and 30), group E (e.g., serotype 4) and group F (e.g., serotypes 40 and 41) (Horwitz et al., supra).

In terms of general structure, all adenoviruses examined to date are nonenveloped, regular icosahedrons of about 65 to 80 nanometers in diameter. Adenoviruses are comprised of linear, double-stranded DNA that is complexed with core proteins and surrounded by the adenoviral capsid. The capsid is comprised of 252 capsomeres, of which 240 are hexons and 12 are pentons. The hexon

WO 98/40509 PCT/US98/05033

3

capsomere provides structure and form to the capsid (Pettersson, in <u>The Adenoviruses</u>, pp. 205-270, Ginsberg, ed., (Plenum Press, New York, NY, 1984)), and is a homotrimer of the hexon protein (Roberts et al., <u>Science</u>, 232, 1148-1151 (1986)). The penton comprises a penton base, which is bound to other hexon capsomeres, and a fiber, which is noncovalently bound to, and projects from, the penton base. The penton fiber protein comprises three identical polypeptides (i.e., polypeptide IV). The Ad2 penton base protein comprises five identical polypeptides (i.e., polypeptides) (i.e., polypeptide III) of 571 amino acids each (Boudin et al., <u>Virology</u>, 92, 125-138 (1979)).

The adenoviruses provide an elegant and efficient means of transferring therapeutic genes into cells. However, one problem encountered with the use of adenoviral vectors for gene transfer in vivo is the generation of antibodies to antigenic epitopes on adenoviral capsid proteins. If sufficient in titer, the antibodies can limit the ability of the vector to be used more than once as an effective gene transfer vehicle. For instance, animal studies demonstrate that intravenous or local administration (e.g., to the lung, heart or peritoneum) of an adenoviral type 2 or 5 gene transfer vector can result in the production of antibodies directed against the vector which prevent expression from the same serotype vector administered 1 to 2 weeks later (see, e.g., Yei et al., supra; Zabner (1994), supra; Setoguchi et al., Am. J. Respir. Cell. Mol. Biol., 10, 369-377 (1994); Kass-Eisler et al., Gene Therapy, 1, 395-402 (1994); Kass-Eisler et al., Gene Therapy 3, 154-162 (1996)). This is a drawback in adenoviral-mediated gene therapy, since many uses of an adenoviral vector (e.g., for prolonged gene therapy) require repeat administration inasmuch as the vector does not stably integrate into the host cell genome. The mechanism by which antibodies

directed against an adenovirus are able to prevent or reduce expression of an adenoviral-encoded gene is unclear. However, the phenomenon is loosely referred to as "neutralization", and the responsible antibodies are termed "neutralizing antibodies."

There are three capsid structures against which neutralizing antibodies potentially can be elicited: fiber, penton, and hexon (Pettersson, supra). The hexon protein, and to a lesser extent the fiber protein, comprise the main antigenic determinants of the virus, and also determine the serotype specificity of the virus (Watson et al., J. Gen. Virol., 69, 525-535 (1988); Wolfort et al., J. Virol., 62, 2321-2328 (1988); Wolfort et al., J. Virol., 56, 896-903 (1985); Crawford-Miksza et al., supra). Researchers have examined and compared the structure of these coat proteins of different adenoviral serotypes in an effort to define the regions of the proteins against which neutralizing antibodies are elicited.

The Ad2 hexon trimer is comprised of a pseudohexagonal base and a triangular top formed of three towers (Roberts et al., <a href="supra">supra</a>; Athappilly et al., <a href="J. Mol.">J. Mol.</a> Biol., 242, 430-455 (1994)). The base pedestal consists of two tightly packed eight-stranded antiparallel beta barrels stabilized by an internal loop. The predominant regions in hexon protein against which neutralizing antibodies are directed appear to be in loops 1 and 2 (i.e., LI or 11, and LII or 12, respectively) in one of the three towers. For instance, Kinloch et al. (J. Biol. Chem., 258, 6431-6436 (1984)) compared adenoviral hexon sequences and theorized that the serotype-specific antigenic determinants on hexon are located in amino acid residues 120 to 470 encompassing the  $\it 11$  and  $\it 12$  loops since type-specific sequence differences are mainly concentrated in this region. Toogood et al. (J. Gen.

<u>Virol.</u>, <u>73</u>, 1429-1435 (1992)) used peptides from this region to generate specific anti-loop antisera and confirmed that antibodies against residues 281-292 of 11 and against residues 441-455 of 12 were sufficient to neutralize infection. Also, Crompton et al. (<u>J. Gen. Virol.</u>, <u>75</u>, 133-139 (1994)) modified these loops to accept neutralizing epitopes from polio virus, and demonstrated that infection with the resultant adenoviral vector generated neutralizing immunity against polio virus. More recently it was demonstrated that the hexon protein is composed of seven discrete hypervariable regions in loops and 1 and 2 (HVR1 to HVR7) which vary in length and sequence between adenoviral serotypes (Crawford-Miksza et al., <u>supra</u>).

Less is known regarding the regions of the fiber protein against which neutralizing antibodies potentially can be directed. However, much data is available on the structure of the fiber protein. The trimeric fiber protein consists of a tail, a shaft, and a knob (Devaux et al., J. Molec. Biol., 215, 567-588 (1990)). The fiber shaft region is comprised of repeating 15 amino acid motifs, which are believed to form two alternating beta strands and beta bends (Green et al., EMBO J., 2, 1357-1365 (1983)). The overall length of the fiber shaft region and the number of 15 amino acid repeats differ between adenoviral serotypes. The receptor binding domain of the fiber protein and sequences necessary for fiber trimerization are localized in the knob region encoded by roughly the last 200 amino acids of the protein (Henry et al., J. Virol., 68(8), 5239-5246 (1994)); Xia et al., Structure, 2(12), 1259-1270 (1994)). Furthermore, all adenovirus serotypes appear to possess a type of specific moiety located in the knob region (Toogood et al., supra.)

Given the existence of these potential epitopes in hexon protein and fiber protein, it is understandable

that, in some cases, difficulties have been encountered using adenovirus as a vector for gene therapy.

Accordingly, recombinant adenoviral vectors capable of escaping such neutralizing antibodies (in the event they are preexisting and hamper gene expression commanded by adenovirus in an initial dose), and which would allow repeat doses of adenoviral vectors to be administered, would significantly advance current gene therapy methodology.

Thus, the present invention seeks to overcome at least some of the aforesaid problems of recombinant adenoviral gene therapy. In particular, it is an object of the present invention to provide a recombinant adenovirus comprising a chimeric coat protein that has a decreased ability or inability to be recognized by antibodies (i.e., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein. These and other objects and advantages of the present invention, as well as additional inventive features, will be apparent from the description of the invention provided herein.

#### BRIEF SUMMARY OF THE INVENTION

The present invention provides a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) comprising a nonnative amino acid sequence. The chimeric adenovirus coat protein is not recognized by, or has a decreased ability to be recognized by, a neutralizing antibody directed against the corresponding wild-type (i.e., native) coat protein. The chimeric adenovirus coat protein enables a vector (such as an adenovirus) comprising the corresponding protein to be administered repetitively, or to be administered following administration of an adenovirus vector comprising the corresponding wild-type coat protein. It also enables a

7

vector (such as an adenovirus) comprising the chimeric protein to be administered and effect gene expression in the case where there are preexisting neutralizing antibodies directed against the wild-type adenovirus coat protein. The present invention also provides a vector, particularly an adenoviral vector, that comprises a chimeric adenovirus coat protein such as chimeric adenovirus hexon protein (and which optionally further comprises a chimeric adenovirus fiber and/or penton base protein), and methods of constructing and using such a vector.

#### BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a diagram of the method employed to construct the vector pAd70-100dlE3.fiber7.

Figure 2 is a partial restriction map of the vector pGBS.59-100(HSF:RGD).

#### DETAILED DESCRIPTION OF THE INVENTION

The present invention provides, among other things, a chimeric adenovirus coat protein. The chimeric adenovirus coat protein comprises a nonnative amino acid sequence, such that the chimeric adenovirus coat protein (or a vector comprising the chimeric adenovirus coat protein) has a decreased ability or inability to be recognized by antibodies (e.g., neutralizing antibodies) directed against the corresponding wild-type adenovirus coat protein.

#### Chimeric Adenovirus Coat Protein

A "coat protein" according to the invention is either an adenoviral penton base protein, an adenoviral hexon protein, or an adenoviral fiber protein. Preferably a coat protein is a adenoviral hexon protein or an adenoviral fiber protein. Any one of the serotypes of

human or nonhuman adenovirus can be used as the source of the coat protein, or its gene or coding sequence. Optimally, however, the adenovirus coat protein is that of a Group B or C adenovirus and, preferably, is that of Ad1, Ad2, Ad3, Ad5, Ad6, Ad7, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, or Ad48.

The chimeric adenovirus coat protein (or a vector, such as adenoviral vector, comprising the chimeric adenovirus coat protein) has a decreased ability or an inability to be recognized by an antibody (e.g., a neutralizing antibody) directed against the corresponding wild-type adenovirus coat protein. A "neutralizing antibody" is an antibody that either is purified from or is present in serum. As used herein, an antibody can be a single antibody or a plurality of antibodies. An antibody is "neutralizing" if it inhibits infectivity of (i.e., cell entry) or gene expression commanded by an adenovirus comprising wild-type coat protein, or if it exerts a substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

An ability or inability of a chimeric coat protein to "be recognized by" (i.e., interact with) a neutralizing antibody directed against the wild-type adenovirus coat protein can be assessed by a variety of means known to those skilled in the art. For instance, the removal of one or more epitopes for a neutralizing antibody present in a wild-type adenovirus coat protein to generate a chimeric adenovirus coat protein will result in a decreased ability or inability of the chimeric coat protein to be recognized by the neutralizing antibody. Also, such a decreased ability or inability to interact with a neutralizing antibody directed against wild-type coat protein can be demonstrated by means of a

neutralization test (see, e.g., Toogood et al., <u>supra;</u> Crawford-Miksza et al., <u>supra;</u> Mastrangeli et al., <u>Human</u> <u>Gene Therapy</u>, <u>7</u>, 79-87 (1996)), or as further described herein.

Generally, an "inability" of a chimeric adenovirus coat protein (or a vector comprising a chimeric adenovirus coat protein) to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein means that such an antibody does not interact with the chimeric coat protein, and/or exhibits no substantial deleterious effect on infectivity of or gene expression commanded by an adenovirus comprising wild-type coat protein, as compared, for instance, to any effect on any other adenoviral property.

A "decreased ability" to be recognized by neutralizing antibody directed against wild-type adenovirus coat protein refers to any decrease in the ability of the chimeric adenovirus coat protein (or a vector comprising the chimeric coat protein) to be recognized by an antibody directed against the corresponding wild-type adenovirus coat protein as compared to the wild-type adenovirus coat protein. such ability/inability is assessed by means of a neutralization test in particular, preferably a "decreased ability" to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein is exhibited by from about a 10% to about a 99% increase in the ability of a recombinant adenovirus comprising the chimeric coat protein to cause a visible cytopathic effect (c.p.e.) in cells such as A549 cells or COS-1 cells (or other such cells appropriate for a neutralization assay) in the presence of the neutralizing antibody as compared to an adenovirus comprising the wild-type coat protein against which the neutralizing antibody is directed.

WO 98/40509 PCT/US98/05033

10

Furthermore, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of cell infectivity by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type protein. Also, a decreased ability or inability of an adenovirus chimeric coat protein (or a vector comprising a chimeric adenovirus coat protein) to interact with a neutralizing antibody can be shown by a reduction of inhibition (from about 10% to about 99%) or no inhibition at all of gene expression commanded by a recombinant vector (such as an adenoviral vector) containing the chimeric coat protein as compared to a recombinant vector containing the wild-type coat protein. These tests can be carried out when the recombinant adenovirus containing the chimeric coat protein is administered following the administration of an adenovirus containing the wild-type coat protein, or when the recombinant adenovirus is administered to a host that has never before encountered or internalized adenovirus (i.e., a "naïve" host). These methods are described, for instance, in the Examples which follow as well as in Mastrangeli et al., supra. Other means such as are known to those skilled in the art also can be employed.

The coat protein is "chimeric" in that it comprises a sequence of amino acid residues that is not typically found in the protein as isolated from, or identified in, wild-type adenovirus, which comprises the so-called native coat protein, or "wild-type coat protein". The chimeric coat protein thus comprises (or has) a "nonnative amino acid sequence" is meant any amino acid sequence (i.e., either component

PCT/US98/05033

11

residues or order thereof) that is not found in the native coat protein of a given serotype of adenovirus, and which preferably is introduced into the coat protein at the level of gene expression (i.e., by production of a nucleic acid sequence that encodes the nonnative amino acid sequence). Generally, the nonnative amino acid sequence can be obtained by deleting a portion of the amino acid sequence, deleting a portion of the amino acid sequence and replacing the deleted amino sequence with a so-called "spacer region", or introducing the spacer region into an unmodified coat protein. Preferably such manipulations result in a chimeric adenovirus coat protein according to the invention that is capable of carrying out the functions of the corresponding wild-type adenovirus coat protein (or, at least that when incorporated into an adenovirus, will allow appropriate virion formation and will not preclude adenoviral-mediated cell entry), and, optimally, that is not impeded in its proper folding. Also, it is desirable that the manipulations do not result in the creation of new epitopes for differing antibodies, unless, of course, such epitopes do not interfere with use of an adenovirus containing the chimeric coat protein as a gene transfer vehicle in vivo.

In particular, a nonnative amino acid sequence according to the invention preferably comprises a deletion of a region of a wild-type adenovirus coat protein, particularly an adenovirus hexon or fiber protein. Optimally the resultant nonnative amino acid sequence is such that one or more of the existing epitopes for neutralizing antibodies directed against the corresponding wild-type adenovirus coat protein have been rendered nonimmunogenic. Desirably, the region deleted comprises from about 1 to about 750 amino acids, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the

region deleted comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. The chimeric coat protein also desirably comprises a plurality of such deletions. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions.

In a preferred embodiment of the present invention, a nonnative amino acid sequence comprises a deletion of one or more regions of a wild-type adenovirus hexon protein, wherein preferably the hexon protein is the Ad2 hexon protein [SEQ ID NO:2] (which is encoded by the sequence of SEQ ID NO:1; GenBank® Data Bank Accession Number U20821), or the Ad5 hexon protein [SEQ ID NO:3] (GenBank® Data Bank Accession Number M73260, which is encoded by the sequence of SEQ ID NO:4), or the Ad7 hexon protein (GenBank® Data Bank Accession Number x76551). Alternately, preferably the hexon protein is the protein sequence reported by Crawford-Miksza et al. (Ad2 hexon [SEQ ID NO:52], Ad5 hexon SEQ ID NO:54]). In particular, the sequences of Crawford-Miksza et al. differ over those reported in the GenBank® Data Bank in that the amino acid residue reported as the first in the Crawford-Miksza et al. sequences is not Met, and the Ad5 hexon sequence is reported as terminating with "Gln His" instead of with "Thr Thr". As employed herein, the numbering of adenovirus hexon amino acid residues corresponds to that in Crawford-Miksza et al.

Desirably the region(s) of the deletion comprises an internal hexon protein sequence ("internal" meaning not at or near the C- or N-terminus of the protein; "near" referring to a distance of 500 amino acids or less), preferably a hypervariable region, e.g., as reported in Crawford-Miksza et al. In particular, optimally, the

internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of 11 loop, preferably from about residue 131 to about residue 331 of the Ad2 hexon protein [SEQ ID NO:6] (which is encoded by the sequence of SEQ ID NO:5), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad5 [SEQ ID NO:8] (which is encoded by the sequence of SEQ ID NO:7), Ad6, Ad7, Ad8, Ad12, Ad16, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more regions (e.g., smaller regions) of the 11 loop. Optimally the region deleted comprises a hypervariable region. Desirably the one or more regions of the 11 loop deleted are regions (i.e., hypervariable regions) selected from this group consisting of the HVR1 region, the HVR2 region, the HVR3 region, the HVR4 region, the HVR5 region, and the HVR6 region. Moreover, preferably the region of the wildtype protein that is deleted (or otherwise manipulated as described herein) occurs on the external surface of the hexon protein. Thus, HVR2, HVR3, HVR4, and HVR5 -- each of which are externally located regions of the hexon . protein -- are particularly preferred for deletion or modification.

The "HVR1 region" preferably comprises from about amino acid 137 to about amino acid 188 of the Ad2 hexon protein [SEQ ID NO:10] (which is encoded by the sequence of SEQ ID NO:9), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:12] (which is encoded by the sequence of SEQ ID NO:11), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48,

BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR2 region" preferably comprises from about amino acid 194 to about amino acid 204 of the Ad2 hexon protein [SEQ ID NO:14] (which is encoded by the sequence of SEQ ID NO:13), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:16] (which is encoded by the sequence of SEQ ID NO:15), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR3 region" preferably comprises from about amino acid 222 to about amino acid 229 of the Ad2 hexon protein [SEQ ID NO:18] (which is encoded by the sequence of SEQ ID NO:17), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:20] (which is encoded by the sequence of SEQ ID NO:19), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR4 region" preferably comprises from about amino acid 258 to about amino acid 271 of the Ad2 hexon protein [SEQ ID NO:22] (which is encoded by the sequence of SEQ ID NO:21), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:24] (which is encoded by the sequence of SEQ ID NO:23), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR5 region" preferably comprises from about amino acid 278 to about amino acid 294 of the Ad2 hexon protein [SEQ ID NO:26] (which is encoded by the sequence

of SEQ ID NO:25), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:28] (which is encoded by the sequence of SEQ ID NO:27), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. In particular, preferably the deleted region comprises from about amino acid 297 to about amino acid 304 just outside of the HVR5 region of the Ad2 hexon protein [SEQ ID NO:30] (which is encoded by the sequence of SEQ ID NO:29), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:32] (which is encoded by the sequence of SEQ ID NO:31), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The "HVR6 region" preferably comprises from about amino acid 316 to about amino acid 327 of the Ad2 hexon protein [SEQ ID NO:34] (which is encoded by the sequence of SEQ ID NO:33), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:36] (which is encoded by the sequence of SEQ ID NO:35), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

In another preferred embodiment of the invention, the internal region of the wild-type hexon protein that is deleted to generate the chimeric hexon protein comprises the entirety of the 12 loop, preferably from about residue 423 to about residue 477 of the Ad2 hexon protein [SEQ ID NO:38] (which is encoded by the sequence of SEQ ID NO:37), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from

Ad1, Ad3, Ad5 [SEQ ID NO:40] (which is encoded by the sequence of SEQ ID NO:39), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. Alternately, preferably the internal region of the wild-type hexon protein that is deleted to produce the chimeric hexon protein comprises one or more smaller regions (e.g., hypervariable regions) of the 12 loop. In particular, preferably the smaller region of the 12 loop comprises the HVR7 region.

The "HVR7 region" preferably comprises from about amino acid 433 to about amino acid 465 of the Ad2 hexon protein [SEQ ID NO:42] (which is encoded by the sequence of SEQ ID NO:41), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:44] (which is encoded by the sequence of SEQ ID NO:43), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra. In particular, preferably the deleted region comprises from about amino acid 460 to about amino acid 466 of the HVR7 region (i.e., extending one base pair outside of this region) of the Ad2 hexon protein [SEQ ID NO:46] (which is encoded by the sequence of SEQ ID NO:45), or the corresponding region from another adenoviral serotype, e.g., particularly the corresponding region from Ad1, Ad3, Ad5 [SEQ ID NO:48] (which is encoded by the sequence of SEQ ID NO:47), Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

Along the same lines, the chimeric adenovirus hexon protein desirably comprises deletions in one or both of the aforementioned regions, i.e., the hexon protein comprises deletions in one or both of the 11 and 12 loops,

which deletions can constitute the entirety of the loop(s), or can comprise deletions of one or more smaller regions (e.g., hypervariable regions) in one or both of the hexon loops. In particular, desirably the deleted region(s) are selected from the group consisting of SEQ ID. NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

An "equivalent" is a naturally occurring variation of an amino acid or nucleic acid sequence, e.g., as are observed among different strains of adenovirus. A conservative variation is a variation of an amino acid sequence that results in one or more conservative amino acid substitution(s). A "conservative amino acid substitution" is an amino acid substituted by an alternative amino acid of similar charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., basic, Arg and Lys; aliphatic Ala, Cys, Gly, Ile, Leu, Met and Val; aromatic, Phe, Tyr, Trp, and His; hydrophilic, Glu, Gln, Asn, and Asp; hydroxyl, Ser and Thr).

In another preferred embodiment, the nonnative amino acid sequence of the chimeric adenoviral coat protein (i.e., particularly a chimeric adenoviral fiber or hexon protein) comprises a deletion of one or more region(s) of the wild-type adenovirus coat protein (particularly the 11)

and/or 12 loops, and, most particularly, the HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and/or HVR7 regions of the wildtype adenovirus hexon protein) as previously described, and further comprises a replacement of the region(s) with a spacer region preferably of from 1 to about 750 amino acids, especially of from about 1 to about 500 amino acids, and particularly of from about 1 to about 300 amino acids. It also is desirable that the region deleted and replaced comprises a smaller region less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. The chimeric coat protein also desirably comprises a plurality of such replacements. Thus, according to the invention, the chimeric adenovirus coat protein comprises modification of one or more amino acids, and such modification is made in one or more regions which can be a smaller region. A spacer region of the aforementioned size also preferably simply can be inserted into one of the aforementioned regions (particularly into the 11 and/or 12 loop, or one or more of the aforementioned HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7 regions of the adenovirus hexon protein) in the absence of any deletion to render the resultant chimeric protein nonimmunogenic by, for instance, destroying the ability of a neutralizing antibody to interact with that particular site (e.g., by changing the spatial juxtaposition of critical amino acids with which the antibody interacts).

Optimally the spacer region comprises a nonconservative variation of the amino acid sequence of wild-type adenovirus coat protein (particularly wild-type adenovirus hexon protein) that comprises an epitope for a neutralizing antibody, and which may or may not be deleted upon the insertion of the spacer region. A "nonconservative variation" is a variation of this amino acid sequence that does not result in the creation or

recreation in the chimeric adenovirus coat protein of the epitope for a neutralizing antibody directed against the wild-type adenovirus coat protein, and, in particular, is a variation of the spacer region that results in one or more nonconservative amino acid insertion(s) or substitution(s) in this region. A "nonconservative amino acid substituted by an alternative amino acid of differing charge density, hydrophilicity/hydrophobicity, size, and/or configuration (e.g., a change of a basic amino acid for an acidic amino acid, a hydrophilic amino acid for a hydrophobic amino acid, and the like).

Desirably the spacer region does not interfere with the functionality of the chimeric adenovirus coat protein, particularly the chimeric adenovirus hexon or fiber protein, e.g., the ability of hexon protein to bind penton base protein or other hexon capsomeres, or the ability of penton fiber to bind penton base and/or to a cell surface receptor. Such functionality can be assessed by virus viability. Similarly, the absence of the creation or recreation of the epitope(s) for a neutralizing antibody directed against the wild-type coat (e.g., hexon and/or fiber) protein can be confirmed using techniques as described in the Examples which follow (e.g., by ensuring the antibody, which may be in a carrier fluid such as serum or other liquid, binds the wild-type adenovirus coat protein, but not the chimeric adenovirus coat protein).

Preferably the spacer region incorporated into the adenovirus coat protein (i.e., either as an insertion into the wild-type coat protein, or to replace one or more deleted region(s) of the wild-type adenovirus coat protein) comprise a series of polar and/or charged amino acids (e.g., Lys, Arg, His, Glu, Asp, and the like), or amino acids with intermediate polarity (e.g., Gln, Asn, Thr, Ser, Met, and the like). In particular, desirably

the spacer region comprises the sequence of SEQ ID NO:50 (which is encoded by the sequence of SEQ ID NO:49), and equivalents and conservative variations of SEQ ID NO:50. Alternately, the spacer region can comprise any other sequence like the FLAG octapeptide sequence of SEQ ID NO:50 that will not interfere with the functionality of the resultant chimeric protein.

In still yet another preferred embodiment, a region of a wild-type adenovirus coat protein (particularly an adenovirus hexon and/or fiber protein) is deleted and replaced with a spacer region comprising the corresponding coat protein region of another adenoviral serotype. Preferably in this embodiment the spacer region is of a different adenoviral group. For instance, preferably a region of an Ad2 coat protein can be replaced with the corresponding region of an Ad5 or Ad7 coat protein (or any other serotype of adenovirus as described above), and vice versa. It also is preferable that such a spacer region comprising the coat protein region of another adenoviral serotype is simply inserted into the corresponding coat protein region of the chimeric coat protein. case, the likelihood of obtaining a chimeric hexon protein that is functional can be increased by making sure that the size of the hypervariable domain resulting from such insertion approximates the size of a known hypervariable domain. For instance, the HVR1 region of Ad40 is about 30 amino acids smaller than the HVR1 region of Ad2 (as well as other adenoviruses such as Ad5, Ad8, etc.). preferably a spacer region of about 30 amino acids can be incorporated into the Ad40 HVR1 region to produce a chimeric adenovirus hexon protein. In particular, desirably the region of Ad2 (or other adenovirus) that is not present in Ad40 (i.e., approximately amino acid residues 138 to 174), or a portion thereof, is introduced

WO 98/40509 PCT/US98/05033

21

into Ad40 to produce the chimeric adenoviral hexon protein.

According to the invention, desirably the nonnative amino acid sequence of a chimeric coat protein comprises a plurality of such replacements or insertions. When the coat protein is incorporated into an adenoviral vector, preferably the entire coat protein of one adenoviral serotype can be substituted with the entire coat protein of another adenoviral serotype, as described further herein.

The region or regions of wild-type adenovirus hexon protein that are deleted and replaced by the spacer region, or into which the spacer region is inserted, can be any suitable region(s) and desirably comprise one or more of the regions described above with respect to the hexon protein deletions. For instance, preferably the one or more regions into which the spacer region is inserted or which the spacer region replaces comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.

Similarly, the spacer region itself (i.e., both for insertion as well as replacement) preferably comprises the entirety of the 11 and/or 12 loop, or a sequence selected

from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48, and equivalents and conservative variations of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:34, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:36, SEQ ID NO:36, SEQ ID NO:46, and SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:46, and SEQ ID NO:48.

The fiber protein also preferably is altered in a similar fashion as described for modification of hexon protein to escape antibodies directed in particular against wild-type adenovirus fiber protein. protein sequences and methods of modifying fiber protein are known to those skilled in the art (see, e.g., Xia et al., supra; Novelli et al., Virology, 185, 365-376 (1991)). The fiber manipulations can be carried out in the absence of, or along with, modifications to the adenovirus hexon protein. In particular, preferably the fiber protein can be replaced in its entirety, or in part, with sequences of a fiber protein from a different serotype of adenovirus. Also, preferably, deletions can be made of fiber sites that constitute an epitope for a neutralizing antibody, and/or insertions can be made at the site to destroy the ability of the protein to interact with the antibody.

# Nucleic Acid Encoding The Chimeric Adenovirus Coat Protein

Preferably the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon or fiber protein) comprises a nonnative amino acid sequence wherein

the alteration is made at the level of DNA. Thus, the invention preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein. Desirably, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of a region (or a plurality of such deletions) that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids. In particular, optimally the deletion (e.g., of an adenoviral hexon protein) comprises the entirety of the 11 and/or 12 loop, or a sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

The invention also preferably provides an isolated and purified nucleic acid encoding a chimeric adenovirus hexon protein as defined herein, wherein the nucleic acid sequence comprises a deletion of one or more sequences selected from the group consisting of equivalents and conservatively modified variants of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID

NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra.

With respect to the nucleic acid sequence, an "equivalent" is a variation on the nucleic acid sequence such as can occur in different strains of adenovirus, and which either does or does not result in a variation at the amino acid level. Failure to result in variation at the amino acid level can be due, for instance, to degeneracy in the triplet code. A "conservatively modified variant" is a variation on the nucleic acid sequence that results in one or more conservative amino acid substitutions. In comparison, a "nonconservatively modified variant" is a variation on the nucleic acid sequence that results in one or more nonconservative amino acid substitutions.

In another preferred embodiment, the invention provides an isolated and purified nucleic acid encoding a chimeric adenovirus coat protein wherein the nucleic acid sequence further comprises a replacement of the deleted region (or a plurality of such replacements) with a spacer nucleic acid region (i.e., the nucleic acid sequence that encodes the aforementioned "spacer region") that encodes from about 1 to about 750 amino acids of the wild-type adenovirus coat protein, preferably from about 1 to about 500 amino acids, and optimally from about 1 to about 300 amino acids. It also is desirable that the region deleted and replaced comprises a smaller region that encodes less than about 200 amino acids, preferably less than about 100 amino acids, and optimally less than about 50 amino acids.

Preferably, the spacer nucleic acid region comprises a FLAG octapeptide-encoding sequence [SEQ ID NO:49], and equivalents and conservatively modified variants of SEQ ID NO:49. Similarly, a spacer nucleic acid region can be employed that substitutes one or more coat protein encoding regions (particularly a hexon protein encoding region) of a particular adenoviral serotype with a coat protein encoding region (particularly a hexon protein encoding region) of another adenoviral serotype. Thus, preferably a spacer nucleic acid region present in a chimeric adenoviral hexon protein is selected from the group consisting of sequences that encode the entirety of the 11 and/or 12 loop, or SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47, or a sequence comprising the corresponding region from Ad1, Ad3, Ad6, Ad7, Ad8, Ad11, Ad12, Ad14, Ad16, Ad21, Ad34, Ad35, Ad40, Ad41, Ad48, BAV3, or MAV1, especially as reported in Crawford-Miksza et al., supra, and equivalents and conservatively modified variants of these sequences.

As described above with respect to the chimeric adenovirus coat protein, the spacer nucleic acid region (or a plurality thereof) simply can be incorporated into the coat protein in the absence of any deletions. These manipulations can be carried out so as to produce the above-described chimeric adenovirus coat protein.

The means of making such a chimeric adenoviral coat protein (i.e., by introducing conservative or nonconservative variations at either the level of DNA or protein) are known in the art, are described in the Examples which follow, and also can be accomplished by means of various commercially available kits and vectors

(e.g., New England Biolabs, Inc., Beverly, MA; Clontech, Palo Alto, CA; Stratagene, LaJolla, CA, and the like). In particular, the ExSite™ PCR-based site-directed mutagenesis kit and the Chameleon™ double-stranded site-directed mutagenesis kit by Stratagene can be employed for introducing such mutations. Moreover, the means of assessing such mutations (e.g., in terms of effect on ability not to be neutralized by antibodies directed against wild-type hexon protein) are described in the Examples herein.

Accordingly, the present invention provides a preferred means of making a chimeric adenoviral coat protein, particularly a chimeric adenoviral hexon protein, which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein (e.g., the wild-type adenovirus hexon protein), and deleting one or more region(s) of the chimeric adenovirus coat protein (particularly the chimeric adenovirus hexon protein) comprising from about 1 to about 750 amino acids by modifying the corresponding nucleic acid coding sequence. Similarly, the invention provides a method of making a chimeric adenovirus coat protein (particularly a chimeric adenovirus hexon protein) which comprises obtaining an adenoviral genome encoding the wild-type adenovirus coat protein, deleting one or more region(s) of the adenovirus coat protein comprising from about 1 to about 750 amino acids by modifying the corresponding coding sequence, and replacing the deleted region(s) with a spacer region comprising from about 1 to about 300 amino acids by introducing a nucleic acid region (i.e., a "spacer nucleic acid region") that codes for same. Alternately, the spacer region preferably is simply incorporated into the coat protein (particularly the hexon protein) in the absence of any deletion. Optimally the spacer nucleic acid region encodes a nonconservative variation of the

WO 98/40509 PCT/US98/05033

amino acid sequence of the wild-type adenovirus coat protein. The size of the DNA used to replace the native coat protein coding sequence may be constrained, for example, by impeded folding of the coat protein or improper assembly of the coat protein into a complex (e.g., penton base/hexon complex) or virion. DNA encoding 150 amino acids or less is particularly preferred for insertion/replacement in the chimeric coat protein gene sequence, and DNA encoding 50 amino acids or less is even more preferred.

Briefly, the method of mutagenesis comprises deleting one or more regions of an adenovirus coat protein, and/or inserting into an adenovirus coat protein one or more regions with a differing amino acid sequence, particularly by manipulating the DNA sequence. Several methods are available for carrying out such manipulations of adenovirus coat protein DNA sequences; these methods further can be used in combination. The method of choice depends on factors known to those skilled in the art, e.g., the size of the DNA region to be manipulated. For instance, convenient restriction sites (which further can be introduced into a sequence) can be used to introduce or remove segments of DNA, or entire genes or coding sequences. Alternately, other methods of mutagenesis involve the hybridization of a mismatched oligonucleotide to a region of single-stranded target DNA, extending the primer, for instance, using T7 DNA polymerase or other such means to produce a double-stranded heteroduplex, and isolating the mutant strand that incorporates the mismatched oligonucleotide from the parental nonmutant strand for use as a template and in further manipulations. The mutant strand can be separated from the parental strand using various selection means known to those skilled in the art (see, e.g., Kunkel et al., Methods Enzymol., 204, 125-139 (1991), as well as the underlying

methodology employed in the Chameleon $^{TM}$  kit). Alternately, the parental strand can be selectively degraded, for instance, with use of enzymes that nick the nonmethylated strand of a hemi-methylated DNA molecule (e.g., HpaII, MspI, and Sau3AI), and by extending the mutant strand using 5-methyl-dCTP, which renders the strand resistant to cleavage by these enzymes. Along the same lines, an entirely PCR-based approach can be employed for making mutations (e.g., Kunkel, Proc. Natl. Acad. Sci., 82, 488-492 (1985); Costa et al., Nucleic Acids Res., 22, 2423 (1994)), for instance, such as the approach encompassed by the  $\mathsf{ExSite}^{\mathsf{TM}}$  kit. More generally, amino acid substitutions or deletions can be introduced during PCR by incorporating appropriate mismatches in one or both primers. Once the chimeric coat protein sequence has been produced, the nucleic acid fragment encoding the sequence further can be isolated, e.g., by PCR amplification using 5' and 3' primers, or through use of convenient restriction sites.

## Vector Comprising a Chimeric Hexon Protein

A "vector" according to the invention is a vehicle for gene transfer as that term is understood by those skilled in the art, and includes viruses, plasmids, and the like. A preferred vector is an adenovirus, particularly a virus of the family Adenoviridae, and desirably of the genus Mastadenovirus (e.g., comprised of mammalian adenoviruses) or Aviadenovirus (e.g., comprised of avian adenoviruses). Such an adenovirus (or other viral vector) can be transferred by its own means of effecting cell entry (e.g., by receptor-mediated endocytosis), or can be transferred to a cell like a plasmid, i.e., in the form of its nucleic acid, for instance, by using liposomes to transfer the nucleic acid, or by microinjecting or transforming the DNA into the cell. The nucleic acid vectors that can be employed for

WO 98/40509 PCT/US98/05033

gene transfer, particularly the adenoviral nucleic acid vectors, are referred to herein as "transfer vectors". Such nucleic acid vectors also include intermediary plasmid vectors that are employed, e.g., in the construction of adenoviral vectors.

Desirably an adenoviral vector is a serotype group C virus, preferably an Ad2 or Ad5 vector, although any other serotype adenoviral vector (e.g., group A including serotypes 12 and 31, group B including serotypes 3 and 7, group D including serotypes 8 and 30, group E including serotype 4, and group F including serotypes 40 and 41, and other Ad vectors previously described) can be employed. An adenoviral vector employed for gene transfer can be replication competent. Alternately, an adenoviral vector can comprise genetic material with at least one modification therein, which renders the virus replication deficient. The modification to the adenoviral genome can include, but is not limited to, addition of a DNA segment, rearrangement of a DNA segment, deletion of a DNA segment, replacement of a DNA segment, or introduction of a DNA lesion. A DNA segment can be as small as one nucleotide and as large as 36 kilobase pairs (i.e., the approximate size of the adenoviral genome) or, alternately, can equal the maximum amount which can be packaged into an adenoviral virion (i.e., about 38 kb). Preferred modifications to the group C adenoviral genome include modifications in the E1, E2, E3 and/or E4 regions. Similarly, an adenoviral vector can be a cointegrate, i.e., a ligation of adenoviral sequences with other sequences, such as other virus sequences, particularly baculovirus sequences, or plasmid sequences, e.g., so as to comprise a prokaryotic or eukaryotic expression vector.

In terms of an adenoviral vector (particularly a replication deficient adenoviral vector), such a vector can comprise either complete capsids (i.e., including a

viral genome such as an adenoviral genome) or empty capsids (i.e., in which a viral genome is lacking, or is degraded, e.g., by physical or chemical means). The capsid further can comprise nucleic acid linked to the surface by means known in the art (e.g., Curiel et al., Human Gene Therapy, 3, 147-154 (1992)) or can transfer non-linked nucleic acid, for instance, by adenoviral-mediated uptake of bystander nucleic acid (e.g., PCT International Application WO 95/21259).

Along the same lines, since methods are available for transferring an adenovirus in the form of its nucleic acid sequence (i.e., DNA), a vector (i.e., a transfer vector) similarly can comprise DNA, in the absence of any associated protein such as capsid protein, and in the absence of any envelope lipid. Inasmuch as techniques are available for making a RNA copy of DNA (e.g., in vitro transcription), and inasmuch as RNA viruses also can be employed as vectors or transfer vectors, a transfer vector also can comprise RNA. Thus, according to the invention whereas a vector comprises (and, further, may encode) a chimeric adenoviral coat protein, a transfer vector typically encodes a chimeric adenoviral coat protein (particularly a chimeric adenoviral hexon and/or fiber protein).

Based on this, the invention provides an adenoviral vector that comprises a chimeric coat protein (particularly a chimeric hexon and/or fiber protein) according to the invention. Preferably such a vector comprises a chimeric coat protein (particularly a chimeric adenovirus hexon protein and/or chimeric adenovirus fiber protein) as described above. Alternately, preferably the vector lacks wild-type fiber protein, e.g., the vector encodes a truncated or non-functional fiber protein, or fails to translate fiber protein. Such fiber mutations and the means of introducing fiber mutations are known to

PCT/US98/05033

those skilled in the art (see, e.g., Falgout et al.,  $\underline{J}$ . Virol., 62, 622-625 (1988)).

Of course, the chimeric adenoviral coat proteins include coat proteins in which the native (i.e., wild-type) hexon and/or fiber protein of an adenoviral vector is replaced by a hexon and or fiber amino acid sequence of a different adenoviral serotype such that the resultant adenoviral vector has a decreased ability or inability to be recognized by neutralizing antibodies directed against the corresponding wild-type coat protein. This replacement can comprise the entirety of the hexon and/or fiber amino acid sequence, or only a portion, as described above. Both proteins can be manipulated (e.g., in a single adenovirus), or only a single chimeric adenovirus coat protein can be employed, with the remaining coat proteins being wild-type.

A vector according to the invention (including a transfer vector) preferably comprises additional sequences and mutations, e.g., some that can occur within the coat protein coding sequence itself. In particular, a vector according to the invention further preferably comprises a nucleic acid encoding a passenger gene or passenger coding sequence. A "nucleic acid" is a polynucleotide (i.e., DNA or RNA). A "gene" is any nucleic acid sequence coding for a protein or an RNA molecule. Whereas a gene comprises coding sequences plus any non-coding sequences, a "coding sequence" does not include any non-coding (e.g., regulatory) DNA. A "passenger gene" or "passenger coding sequence" is any gene which is not typically present in and is subcloned into a vector (e.g., a transfer vector) according to the present invention, and which upon introduction into a host cell is accompanied by a discernible change in the intracellular environment (e.g., by an increased level of deoxyribonucleic acid (DNA), ribonucleic acid (RNA), peptide or protein, or by an

altered rate of production or degradation thereof). A "gene product" is either an as yet untranslated RNA molecule transcribed from a given gene or coding sequence (e.g., mRNA or antisense RNA) or the polypeptide chain (i.e., protein or peptide) translated from the mRNA molecule transcribed from the given gene or coding sequence. A gene or coding sequence is "recombinant" if the sequence of bases along the molecule has been altered from the sequence in which the gene or coding sequence is typically found in nature, or if the sequence of bases is not typically found in nature. According to this invention, a gene or coding sequence can be naturally occurring or wholly or partially synthetically made, can comprise genomic or complementary DNA (cDNA) sequences, and can be provided in the form of either DNA or RNA.

Non-coding sequences or regulatory sequences include promoter sequences. A "promoter" is a DNA sequence that directs the binding of RNA polymerase and thereby promotes RNA synthesis. "Enhancers" are cis-acting elements of DNA that stimulate or inhibit transcription of adjacent genes. An enhancer that inhibits transcription is also termed a "silencer". Enhancers differ from DNA-binding sites for sequence-specific DNA binding proteins found only in the promoter (which also are termed "promoter elements") in that enhancers can function in either orientation, and over distances of up to several kilobase pairs, even from a position downstream of a transcribed region. According to the invention, a coding sequence is "operably linked" to a promoter (e.g., when both the coding sequence and the promoter constitute a passenger gene) when the promoter is capable of directing transcription of that coding sequence.

Accordingly, a "passenger gene" can be any gene, and desirably either is a therapeutic gene or a reporter gene. Preferably a passenger gene is capable of being expressed

in a cell in which the vector has been internalized. For instance, the passenger gene can comprise a reporter gene, or a nucleic acid sequence which encodes a protein that can be detected in a cell in some fashion. The passenger gene also can comprise a therapeutic gene, for instance, a therapeutic gene which exerts its effect at the level of RNA or protein. Similarly, a protein encoded by a transferred therapeutic gene can be employed in the treatment of an inherited disease, such as, e.g., the cystic fibrosis transmembrane conductance regulator cDNA for the treatment of cystic fibrosis. The protein encoded by the therapeutic gene can exert its therapeutic effect by resulting in cell killing. For instance, expression of the gene in itself may lead to cell killing, as with expression of the diphtheria toxin A gene, or the expression of the gene may render cells selectively sensitive to the killing action of certain drugs, e.g., expression of the HSV thymidine kinase gene renders cells sensitive to antiviral compounds including acyclovir, gancyclovir and FIAU (1-(2-deoxy-2-fluoro-b-Darabinofuranosil)-5-iodouracil). Moreover, the therapeutic gene can exert its effect at the level of RNA, for instance, by encoding an antisense message or ribozyme, by affecting splicing or 3' processing (e.g., polyadenylation), or by encoding a protein which acts by affecting the level of expression of another gene within the cell (i.e., where gene expression is broadly considered to include all steps from initiation of transcription through production of a processed protein), perhaps, among other things, by mediating an altered rate of mRNA accumulation, an alteration of mRNA transport, and/or a change in post-transcriptional regulation. Accordingly, the use of the term "therapeutic gene" is intended to encompass these and any other embodiments of that which is more commonly referred to as gene therapy

and is known to those of skill in the art. Similarly, the recombinant adenovirus can be used for gene therapy or to study the effects of expression of the gene (e.g., a reporter gene) in a given cell or tissue in vitro or in vivo, or for diagnostic purposes.

Also, a passenger coding sequence can be employed in the vector. Such a coding sequence can be employed for a variety of purposes even though a functional gene product may not be translated from the vector sequence. For instance, the coding sequence can be used as a substrate for a recombination reaction, e.g., to recombine the sequence with the host cell genome or a vector resident in the cell. The coding sequence also can be an "anticoding sequence," e.g., as appropriate for antisense approaches. Other means of using the coding sequence will be known to one skilled in the art.

The present invention thus provides recombinant adenoviruses comprising a chimeric hexon protein and/or a chimeric fiber protein, and which preferably additionally comprise a passenger gene or genes capable of being expressed in a particular cell. The recombinant adenoviruses can be generated by use of a vector, specifically, a transfer vector, and preferably a viral (especially an adenoviral) or plasmid transfer vector, in accordance with the present invention. Such a transfer vector preferably comprises a chimeric adenoviral hexon and/or fiber gene sequence as previously described.

Similarly, the means of constructing such a transfer vector are known to those skilled in the art. For instance, a chimeric adenovirus coat protein gene sequence can simply be ligated into the vector using convenient restriction sites. Alternately, a wild-type adenovirus gene sequence can be mutagenized to create the chimeric coat protein sequence following its subcloning into a vector. Similarly, a chimeric coat protein gene sequence

can be moved via standard molecular genetic techniques from a transfer vector into baculovirus or a suitable prokaryotic or eukaryotic expression vector (e.g., a viral or plasmid vector) for expression and evaluation of penton base binding, and other biochemical characteristics.

Accordingly, the present invention also provides recombinant baculoviral and prokaryotic and eukaryotic expression vectors comprising an aforementioned chimeric adenoviral coat protein gene sequence, which, along with the nucleic acid form of the adenoviral vector (i.e., an adenoviral transfer vector) are "transfer vectors" as defined herein. By moving the chimeric gene from an adenoviral vector to baculovirus or a prokaryotic or eukaryotic expression vector, high protein expression is achievable (approximately 5-50% of the total protein being the chimeric protein).

Similarly, adenoviral vectors (e.g., virions or virus particles) are produced using transfer vectors. For instance, an adenoviral vector comprising a chimeric coat protein according to the invention can be constructed by introducing into a cell, e.g., a 293 cell, a vector comprising sequences from the adenoviral left arm, and a vector comprising sequences from the adenoviral right arm, wherein there is a region of overlap between the sequences. As described in the Examples which follow, this methodology results in recombination between the sequences, generating a vector that comprises a portion of each of the vectors, particularly the region comprising the chimeric coat protein sequences.

The present invention thus preferably also provides a method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus hexon protein and/or fiber protein. This method comprises replacing a coat protein of the vector

(i.e., a wild-type adenovirus hexon and/or fiber protein) with the corresponding chimeric adenovirus coat protein according to the invention to produce a recombinant adenoviral vector.

The coat protein chimera-containing particles are produced in standard cell lines, e.g., those currently used for adenoviral vectors. Deletion mutants lacking the fiber gene, or possessing shortened versions of the fiber protein, similarly can be employed in vector construction, e.g., H2d1802, H2d1807, H2d1021 (Falgout et al., supra), as can other fiber mutants. The fiberless particles have been shown to be stable and capable of binding and infecting cells (Falgout et al., supra).

# Illustrative Uses and Benefits

The present invention provides a chimeric coat protein that has a decreased ability or inability to be recognized by a neutralizing antibody directed against the corresponding wild-type coat protein, as well as vectors (including transfer vectors) comprising same. The chimeric coat protein (such as a chimeric hexon and/or fiber protein) has multiple uses, e.g., as a tool for studies in vitro of capsid structure and assembly, and capsomere binding to other proteins.

A vector (e.g., a transfer vector) comprising a chimeric coat protein can be used in strain generation, for instance, in generation of recombinant strains of adenovirus. Similarly, such a vector, particularly an adenoviral vector, can be used in gene therapy. Specifically, a vector of the present invention can be used to treat any one of a number of diseases by delivering to targeted cells corrective DNA, i.e., DNA encoding a function that is either absent or impaired, or a discrete killing agent, e.g., DNA encoding a cytotoxin that, for instance, is active only intracellularly.

Diseases that are candidates for such treatment include, but are not limited to, cancer, e.g., melanoma, glioma or lung cancers; genetic disorders, e.g., cystic fibrosis, hemophilia or muscular dystrophy; pathogenic infections, e.g., human immunodeficiency virus, tuberculosis or hepatitis; heart disease, e.g., preventing restenosis following angioplasty or promoting angiogenesis to reperfuse necrotic tissue; and autoimmune disorders, e.g., Crohn's disease, colitis or rheumatoid arthritis. In particular, gene therapy can be carried out in the treatment of diseases, disorders, or conditions that require repeat administration of the corrective DNA and/or the adenoviral vector, and thus for which current adenoviral-mediated approaches to gene therapy are less than optimal.

Moreover, such a vector, particularly an adenoviral vector, can be used to deliver material to a cell not as a method of gene therapy, but for diagnostic or research purposes. In particular, a vector comprising a chimeric adenovirus coat protein according to the invention can be employed to deliver a gene either in vitro or in vivo, for research and/or diagnostic purposes.

For instance, instead of transferring a so-called therapeutic gene, a reporter gene or some type of marker gene can be transferred instead. Marker genes and reporter genes are of use, for instance, in cell differentiation and cell fate studies, as well as potentially for diagnostic purposes. Moreover, a standard reporter gene such as a  $\beta$ -galactosidase reporter gene, a gene encoding green fluorescent protein (GFP), or a  $\beta$ -glucuronidase gene can be used in vivo, e.g., as a means of assay in a living host, or, for instance, as a means of targeted cell ablation (see, e.g., Minden et al., BioTechniques, 20, 122-129 (1996); Youvan, Science, 268,

264 (1995); U.S. Patent 5,432,081; Deonarain et al., <u>Br.</u>
<u>J. Cancer</u>, 70, 786-794 (1994)).

Similarly, it may be desirable to transfer a gene to use a host essentially as a means of production in vivo of a particular protein. Along these lines, transgenic animals have been employed, for instance, for the production of recombinant polypeptides in the milk of transgenic bovine species (e.g., PCT International Application WO 93/25567). The use of an adenovirus according to the invention for gene transfer conducted for protein production in vivo further is advantageous in that such use should result in a reduced (if not absent) immune response as compared with the use of a wild-typeadenovirus vector. Other "non-therapeutic" reasons for gene transfer include the study of human diseases using an animal model (e.g., use of transgenic mice and other transgenic animals including p53 tumor suppressor gene knockouts for tumorigenic studies, use of a transgenic model for impaired glucose tolerance and human Alzheimer's amyloid precursor protein models for the study of glucose metabolism and for the pathogenesis of Alzheimer's disease, respectively, etc.).

Furthermore, an adenoviral vector comprising a chimeric adenovirus coat protein and employed as described above is advantageous in that it can be isolated and purified by conventional means. For instance, it is likely that special cell lines will not need to be made in order to propagate adenoviruses comprising the chimeric coat proteins.

These aforementioned illustrative uses and recitation of benefits are by no means comprehensive, and it is intended that the present invention encompass such further uses which necessarily flow from, but are not explicitly recited, in the disclosure herein.

# Means of Administration

The vectors and transfer vectors of the present invention can be employed to contact cells either in vitro or in vivo. According to the invention "contacting" comprises any means by which a vector is introduced intracellularly; the method is not dependent on any particular means of introduction and is not to be so construed. Means of introduction are well known to those skilled in the art, and also are exemplified herein.

Accordingly, introduction can be effected, for instance, either in vitro (e.g., in an ex vivo type method of gene therapy or in tissue culture studies) or in vivo by methods that include, but are not limited to, electroporation, transformation, transduction, conjugation, triparental mating, (co-)transfection, (co-) infection, high velocity bombardment with DNA-coated microprojectiles, incubation with calcium phosphate-DNA precipitate, direct microinjection into single cells, and the like. Similarly, the vectors can be introduced by means of membrane fusion using cationic lipids, e.g., liposomes. Such liposomes are commercially available (e.g., Lipofectin®, Lipofectamine™, and the like, supplied by Life Technologies, Gibco BRL, Gaithersburg, MD). Moreover, liposomes having increased transfer capacity and/or reduced toxicity in vivo (see, e.g., PCT International Application WO 95/21259 and references reviewed therein) can be employed in the present invention. Other methods also are available and are known to those skilled in the art.

According to the invention, a "host" encompasses any host into which a vector of the invention can be introduced, and thus encompasses an animal, including, but not limited to, an amphibian, bird, insect, reptile, or mammal. Optimally a host is a mammal, for instance, a

rodent, primate (such as chimpanzee, monkey, ape, gorilla, orangutan, or gibbon), feline, canine, ungulate (such as ruminant or swine), as well as, in particular, a human.

Similarly, a "cell" encompasses any cell (or collection of cells) from a host into which an adenoviral vector can be introduced, e.g., preferably an epithelial cell. Any suitable organs or tissues or component cells can be targeted for vector delivery. Preferably, the organs/tissues/cells employed are of the circulatory system (e.g., heart, blood vessels or blood), respiratory system (e.g., nose, pharynx, larynx, trachea, bronchi, bronchioles, lungs), gastrointestinal system (e.g., mouth, pharynx, esophagus, stomach, intestines, salivary glands, pancreas, liver, gallbladder), urinary system (e.g., kidneys, ureters, urinary bladder, urethra), nervous system (e.g. brain and spinal cord, or special sense organs such as the eye) and integumentary system (e.g., skin). Even more preferably the cells being targeted are selected from the group consisting of heart, blood vessel, lung, liver, gallbladder, urinary bladder, and eye cells.

Thus, the present invention preferably also provides a method of genetically modifying a cell. This method preferably comprises contacting a cell with a vector comprising a chimeric adenovirus hexon protein and/or a chimeric adenovirus fiber protein, wherein desirably the vector is an adenovirus vector. The method preferably results in the production of a host cell comprising a vector according to the invention.

Moreover, the method of the invention of genetically modifying a cell can be employed in gene therapy, or for administration for diagnosis or study. The application of this method in vivo optimally comprises administering to a patient in need of gene therapy (e.g., a patient suffering from a disease, condition or disorder) a therapeutically effective amount of a recombinant adenovirus vector

WO 98/40509 PCT/US98/05033

41

according to the invention. This method preferably can be employed as part of an ongoing gene therapy regimen, e.g., wherein the vector (e.g., a recombinant adenovirus vector) comprising the chimeric adenovirus coat protein is administered following (e.g., after from about 1 week to about 2 months) administration of a therapeutically effective amount of a vector comprising either the corresponding wild-type coat protein or a coat protein of a different adenoviral serotype. Alternately, the vector comprising the chimeric adenovirus coat protein can be employed as an initial attempt at gene delivery.

One skilled in the art will appreciate that suitable methods of administering a vector (particularly an adenoviral vector) of the present invention to an animal for purposes of gene therapy (see, for example, Rosenfeld et al. (1991), supra; Jaffe et al., Clin. Res., 39(2), 302A (1991); Rosenfeld et al., Clin. Res., 39(2), 311A (1991a); Berkner, supra), chemotherapy, vaccination, diagnosis, and/or further study are available. Although more than one route can be used for administration; a particular route can provide a more immediate and more effective reaction than another route. For instance, local or systemic delivery can be accomplished by administration comprising application or instillation of the formulation into body cavities, inhalation or insufflation of an aerosol, or by parenteral introduction, comprising intramuscular, intravenous, peritoneal, subcutaneous, intradermal, as well as topical administration. Clinical trials regarding use of gene therapy vectors in vivo are ongoing. The methodology employed for such clinical trials as well as further technologies known to those skilled in the art can be used to administer the vector of the present invention for the purpose of research, diagnosis and/or gene therapy.

Pharmaceutically acceptable excipients also are well-known to those who are skilled in the art, and are readily available. The choice of excipient will be determined in part by the particular method used to administer the recombinant vector. Accordingly, there is a wide variety of suitable formulations for use in the context of the present invention. The following methods and excipients are merely exemplary and are in no way limiting.

Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the compound dissolved in diluents, such as water, saline, or orange juice; (b) capsules, sachets or tablets, each containing a predetermined amount of the active ingredient, as solids or granules; (c) suspensions in an appropriate liquid; and (d) suitable emulsions. Tablet forms can include one or more of lactose, mannitol, corn starch, potato starch, microcrystalline cellulose, acacia, gelatin, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, stearic acid, and other excipients, colorants, diluents, buffering agents, moistening agents, preservatives, flavoring agents, and pharmacologically compatible excipients. Lozenge forms can comprise the active ingredient in a flavor, usually sucrose and acacia or tragacanth, as well as pastilles comprising the active ingredient in an inert base, such as gelatin and glycerin, or sucrose and acacia, emulsions, gels, and the like containing, in addition to the active ingredient, such excipients as are known in the art.

A vector of the present invention (including an adenoviral vector and a transfer vector), alone or in combination with other suitable components, can be made into aerosol formulations to be administered via inhalation. These aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like.

They may also be formulated as pharmaceuticals for non-pressured preparations such as in a nebulizer or an atomizer.

Formulations suitable for parenteral administration include aqueous and non-aqueous, isotonic sterile injection solutions, which can contain anti-oxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. formulations can be presented in unit-dose or multi-dose sealed containers, such as ampules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described.

Additionally, a vector of the present invention can be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases.

Formulations suitable for vaginal administration can be presented as pessaries, tampons, creams, gels, pastes, foams, or spray formulas containing, in addition to the active ingredient, such carriers as are known in the art to be appropriate.

The dose administered to an animal, particularly a human, in the context of the present invention will vary with the gene of interest, the composition employed, the method of administration, the particular site and organism undergoing administration, and the reason for the administration (e.g., gene therapy, diagnosis, means of producing a protein, further study, etc). Generally, the "effective amount" of the composition is such as to

produce the desired effect in a host which can be monitored using several end-points known to those skilled in the art. For example, one desired effect might comprise effective nucleic acid transfer to a host cell. Such transfer can be monitored in terms of a therapeutic effect (e.g., alleviation of some symptom associated with the disease or syndrome being treated), or by further evidence of the transferred gene or coding sequence or its expression within the host (e.g., using the polymerase chain reaction, Northern or Southern hybridizations, or transcription assays to detect the nucleic acid in host cells, or using immunoblot analysis, antibody-mediated detection, or particularized assays to detect protein or polypeptide encoded by the transferred nucleic acid, or impacted in level or function due to such transfer). One such particularized assay described in the Examples which follow includes an assay for expression of a chloramphenicol acetyl transferase reporter gene.

Generally, to ensure effective transfer of the vectors of the present invention, it is preferable that from about 1 to about 5,000 copies of the vector be employed per cell to be contacted, based on an approximate number of cells to be contacted in view of the given route of administration. It is even more preferable that from about 1 to about 300 plaque forming units (pfu) enter each cell. However, this is just a general guideline which by no means precludes use of a higher or lower amount of a component, as might be warranted in a particular application, either in vitro or in vivo. For example, the actual dose and schedule can vary depending on whether the composition is administered in combination with other pharmaceutical compositions, or depending on interindividual differences in pharmacokinetics, drug disposition, and metabolism. Similarly, amounts can vary in in vitro applications depending on the particular cell

type utilized or the means by which the vector is transferred. One skilled in the art easily can make any necessary adjustments in accordance with the necessities of the particular situation.

The following examples further illustrate the present invention and, of course, should not be construed as in any way limiting its scope.

# Example 1

This example describes experiments investigating adenoviral anti-vector neutralizing immunity.

To clarify the phenomenon of neutralizing immunity, an animal having circulating antibodies to one adenoviral vector type received intratracheal administration of another serotype adenoviral vector, and gene expression commanded by the second vector was monitored.

Specifically, either an Ad4 or Ad5 wild-type vector was administered to the lungs of Sprague-Dawley rats. Ten days later, an Ad5 reporter vector was administered to the lungs of the same animals. This reporter vector, which is referred to herein as the "pure 5" vector, comprises an El-E3- type 5 adenoviral vector which expresses the chloramphenical acetyl transferase (CAT) gene driven by the cytomegalovirus early/intermediate promoter/enhancer (CMV) (i.e., AdCMVCATgD described in Kass-Eisler et al., Proc. Natl. Acad. Sci., 15, 11498-11502 (1993)).

About twenty-four hours following administration of the "pure 5" vector, CAT activity was measured in homogenized lung tissue using a CAT assay as previously described (Kass Eisler et al. (1993), <a href="mailto:supra">supra</a>). CAT activity was monitored at various times thereafter up to 10 days following introduction of the "pure 5" vector. CAT activity was determined relative to the "pure 5" vector administered to naive animals (i.e., expression measured under this condition was considered 100%). The

results of these studies are set out in **Table 1**, and are further reported in Mastrangeli et al., <u>Human Gene</u>
Therapy, 1, 79-87 (1996).

Table 1. Effect of anti-serotype 4 (group E)
neutralizing antibodies on the ability of a "pure 5"
adenoviral vector to transfer a CAT reporter gene to
the lung

Time (0 hours)	Time (10 days)	CAT Activity
		0%
	pure 5	100%
Ad5	pure 5	0%
Ad4	pure 5	105±10%

These results confirm that in the presence of neutralizing antibodies elicited against one adenoviral group (e.g., against group E, serotype 4), it is possible to efficiently transfer and express a gene in vivo using an adenoviral vector derived from another group (e.g., derived from group C, serotype 5). Neutralizing immunity evoked against one serotype group does not protect against infection by another group of adenovirus. These data support the paradigm of alternating adenoviral vectors derived from different subgroups as a strategy to circumvent anti-adenoviral humoral immunity.

# Example 2

The predominant epitopes that evoke neutralizing immunity are located on the fiber and hexon, but mainly on hexon. Based on this, the effect of switching the fiber protein was investigated. A vector was constructed that was identical to the "pure 5" vector except that the fiber gene was switched from a serotype 5, group C fiber to a

serotype 7, group B fiber. The resultant vector is referred to herein as the "5 base/7 fiber" vector.

The Ad5/Ad7 fiber construct was generated as shown in Figure 1. An approximately 2.7 kb (Ad5 28689-31317 bp) fragment in pAd70-100 was replaced with a PacI linker (pAd70-100dlE3.Pac). A BamHI linker was inserted at a MunI site as indicated in Figure 2 to produce pAd70-100dlE3.Pac.Bam. A PCR-amplified PacI-BamHI fragment of approximately 1.1 kb containing the Ad7 fiber gene was inserted into pAd70-100dlE3.Pac.Bam to produce pAd70-100dlE3.fiber7.

In order to assess the ability of the Ad5 virus with Ad7 fiber to infect cells in vitro and in vivo, reporter gene assays were performed. A replication-defective recombinant adenoviral reporter vector designated AdCMV-CATNeo was used in the reporter gene assay. The reporter vector consists of the adenoviral origin of replication and viral packaging sequences, a combination of strong eukaryotic promoter (cytomegalovirus or CMV-1) and splicing elements, the bacterial chloramphenical acetyl transferase (CAT) gene sequence, the mouse  $\beta^{\text{maj}}$ -globin poly(A) site, the neomycin gene sequence (Neo), and sufficient adenoviral DNA to allow for overlap recombination.

The reporter vector was used to generate AdCMV-CATNeo, AdCMV-CATNeo-dlE3 (AdCMV-CATNeo + pAd70-100dlE3) and AdCMV-CATNeo-dlE3-Fiber7 (AdCMV-CATNeo + pAd70-1001E3.Fiber7) viruses. Each virus was grown in large scale, i.e., a one liter suspension of human embryonic kidney 293 cells, to yield virus at a concentration of 10<sup>12</sup> particles/ml. A549 cells were infected with an estimated 100, 300 or 1,000 particles/cell of one of the three viruses. After 48 hours, the cells were harvested and lysates were prepared as described in Kass-Eisler et al.

(1993), supra. Using 50  $\mu$ l of each lysate, CAT assays were performed and acetylated chloramphenical products were separated by thin layer chromatography using chloroform:methanol (95:5). The results of the assays confirm that each virus was able to infect cells and express gene products at appropriate levels. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could infect cells and express genes like the parental virus.

Following this study, adult Sprague-Dawley rats were infected with 10<sup>8</sup> viral particles by direct cardiac injection as described in Kass-Eisler et al. (1993), supra. Five days later, the rats were sacrificed, cardiac lysates were prepared, and CAT assays were performed. amount of the CAT gene product produced was compared between the dlE3 and dlE3-Fiber7 viruses. Results indicated that both viruses were able to infect cells in The replacement of the wild-type Ad5 fiber gene with that of Ad7 did not impair the ability of the virus to infect cells. Accordingly, the virus in which the native fiber was replaced with a nonnative fiber could also infect cells and express genes like the parental virus in vivo. These results support the utility of adenovirus with chimeric fiber in the context of gene therapy.

# Example 3

This example describes the effect on neutralizing immunity of switching the fiber protein of an adenovirus from one serotype to another.

The "pure 5" and "5 base/7 fiber" vectors described in the preceding Example were administered to Sprague-Dawley rats which either were naive or pre-immunized against wild-type Ad5. For these experiments, wild-type Ad5 or wild-type Ad7 (6 x 10 particles in phosphate

WO 98/40509 PCT/US98/05033

buffered saline (PBS)) was administered intraperitoneally as a primary inoculation. Seventeen days later, serum samples were taken, and about 6 x 109 particles in about 50 µl of PBS was injected. At about 120 hours following injection the animals were sacrificed, serum and heart tissue were harvested, and heart tissue was processed for CAT assays as previously described (Kass-Eisler et al. (1993), supra). CAT assays also were performed on heart lysates of rat hearts infected with the "pure 5" vector or "5 base/7 fiber" vector alone.

Administration of either vector to naive animals resulted in comparable levels of CAT in heart tissue. In comparison, administration of either the "pure 5" vector or the "5 base/7 fiber" vector to the animals that were pre-immunized against the "pure 5" vector resulted in a reduction of CAT levels by more than two orders of magnitude as compared with mock-infected controls. These and further results are reported in Gall et al., <u>J.</u> Virol., 70, 2116-2163 (1996).

These results confirm that switching the fiber from that of adenoviral serotype 5 group C vector to that of an adenoviral serotype 7 group B vector by itself is insufficient to allow the vector to escape neutralizing antibodies generated against an adenoviral vector comprising Ad5 fiber. These results imply that antibodies against adenoviral structures other than fiber also are important in the process of neutralizing immunity. Furthermore, whereas switching the fiber serotype to another serotype may be insufficient in and of itself to allow an adenovirus to escape immune detection, such switching when done in combination with removal of other epitopes may be desirable, for instance, to reduce an immune response.

# Example 4

This example describes the construction of adenovirus vectors wherein the neutralizing immunity-evoking epitopes have been modified. In particular, this example describes vectors comprising chimeric adenoviral hexon protein, wherein the hexon neutralizing immunity-evoking epitopes are modified.

The results of the prior example indicate that it is possible to develop vectors for repeat administration in gene therapy from non-group C adenovirus, thus circumventing pre-existing neutralizing immunity. As another strategy, the dominant neutralizing immunity-evoking epitopes on existing group C vectors can be modified to render the vectors less susceptible (or "stealth") to the existing neutralizing immunity. For instance, adenoviral type 5-based E1 E3 CAT-expressing vectors can be constructed that have the same genetic composition as the "pure 5" and "5 base/7 fiber" vectors described above, except for possessing a gene encoding a chimeric hexon that is not recognized by pre-existing anti-type 5 neutralizing immunity.

To derive the vectors, the chimeric hexon gene present in the "pure 5" parental vector can be modified, in particular, 11 and/or 12 can be altered. The hexon modifications that can be made on the "pure 5" CAT vector, or other adenoviral vector (such as any other adenoviral serotype vector), include, but are not limited to: (1) hexon with 11 deleted in its entirety; (2) hexon with 12 deleted in its entirety; (3) hexon with both 11 and 12 deleted; (4) hexon with any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7, deleted; (5)-(8) hexon with a FLAG octamer epitope (i.e., Asp Tyr Lys Asp Asp Asp Asp Lys [SEQ ID NO:50]; Hopp et al., Biotechnology, 6, 1205-1210 (1988)) substituted for 11, 12, or both 11 and 12, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5,

HVR6 or HVR7; (9)-(12) hexon with a FLAG octamer epitope [SEQ ID NO:50] inserted into 11, 12, or both 11 and 12; (13)-(16) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, substituted for 11, 12, both 11 and 12, respectively, or for any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; (17)-(20) hexon with comparable epitopes from Ad7 (group B) (GenBank® Data Bank Accession Number x76551 for Ad7 hexon, and Number M73260 for Ad5 hexon) or Ad2, or any other adenoviral serotype, inserted into 11, 12, both 11 and 12, respectively, or any one or more of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7; and (21) complete substitution of the hexon from Ad2 or another adenoviral serotype, for the Ad5 hexon. The use of the FLAG octamer epitope provides a sequence for incorporation in the chimeric hexon protein that is different from the Ad5 hexon loop sequences, and also provides a positive control using available specific anti-FLAG antibodies (Hopp et al., supra).

These chimeric hexon proteins (and vectors containing them) can be made in several steps. To modify the hexon in the "pure 5" vector, a viral or plasmid vector can be constructed to contain the hexon type 5 coding sequence in a cassette that can be easily modified. The hexon is read off the 1 strand of the L3 transcription unit, i.e., map units 51.6 to 59.7, comprising a region of about 2.9 kb. The two other transcripts that also are encoded by L3 -- i.e., polypeptide VI and a 23 kDa protein -- do not overlap the hexon coding sequence. Moreover, there are no other coding sequences on the r strand that would be altered by the modification of the hexon coding sequence.

Thus, all the modifications of the type 5 hexon can be made using a "hexon 5 cassette" comprised of an

approximate 6.7 kb <u>SfiI-SfiI</u> fragment of the "pure 5" CAT vector. SfiI cuts Ad5 into 3 fragments, the center 6.7 kb fragment (i.e., comprising about 16,282 to 22,992 base pairs, as identified by agarose gel electrophoresis) of which contains all of the L3 region plus some overlap. The "hexon 5 cassette" can be subcloned into a commercially available vector having restriction sites and the like making the vector easily manipulable in terms of modification and recovery of subcloned sequences. One such vector appropriate for subcloning is either the SK or KS version of the pBlueScript® phagemid (Stratagene, LaJolla, CA).

The "hexon 5 cassette" can be mutagenized to generate site-specific mutations in the cloned DNA segment. Several methods are available for carrying out sitespecific mutagenesis. The 11 and 12 deletions, insertions, or replacements (or deletions, insertions, or replacements in HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, or HVR7 regions contained therein) can be made by deleting the relevant sequences using restriction enzymes that cut uniquely within the vector inserts, or other similar means, e.g., by ligating in an end-polished, or otherwise modified, PCR product. Alternately, the hexon sequence contained in the hexon 5 cassette can be modified, e.g., using single-stranded mutagenesis in M13mp8 or some other convenient vector, and using appropriate oligonucleotides encompassing the flanking sequences for identification of plaques as described by Crompton et al., supra. Alternately, a commercially available kit such as the  $\texttt{ExSite}^{\texttt{TM}} \ \texttt{PCR-based} \ \texttt{site-directed} \ \texttt{mutagenesis} \ \texttt{kit} \ \texttt{and} \ \texttt{the}$  $\texttt{Chameleon}^{\texttt{TM}} \ \texttt{double-stranded} \ \texttt{site-directed} \ \texttt{mutagenesis} \ \texttt{kit}$ by Stratagene can be used to introduce insertions, point mutations, or deletions into the chimeric hexon sequence without any need for subcloning into an M13, or other special vector.

Similarly, the FLAG octapeptide sequence (Hopp et al., supra) can be introduced into the vectors (i.e., in the presence or absence of any deletion) by inserting the relevant 24 base pair sequence (GAY TAY AAR GAY GAY GAY GAY AAR [SEQ ID NO:50], wherein Y is C or T/U, and R is A or G)). The replacement of Ad5 hexon loop epitopes with comparable sequences of Ad7, Ad2, or any other adenoviral serotype, or an incorporation of these sequences in the absence of any deletion, can be accomplished by using unique restriction sites, or using one of the aforementioned means of mutagenesis. This usefully creates new serotypes of adenoviral vectors. For example, The replacement of the wildtype hexon protein of Ad5 with the chimeric Ad5 hexon comprising Ad7 hexon loops 1 and 2 gives rise to an adenoviral vector that is effectively neutralized by Ad7 neutralizing antibodies (i.e., neutralizing antibodies raised in response to Ad7 innoculation of a naïve animal), but not by Ad5 neutralizing antibodies.

Moreover, both hypervariable loops 1 and 2 can be deleted from a serotype 5 or another serotype adenoviral vector. Adenoviral vectors and there genomes comprising these deletions are useful as a starting point to create other adenoviral vectors having loop replacements, as a tool for studying hexon structure-function relationships, and under some circumstances as a gene transfer vector with limited vulnerability to the adaptive immune system.

### Example 5

This example describes the method of replacing the hexon protein of one serotype adenoviral vector with the hexon protein of another serotype adenoviral vector to generate a recombinant adenovirus. As representative of this method, the hexon protein of an Ad5 vector was replaced with the hexon protein of an Ad2 vector. This

example also describes the method of incorporating the chimeric hexon proteins of the preceding Example into a vector to make a recombinant adenovirus.

Using standard molecular biology techniques, the Ad5 hexon gene open reading frame (ORF) was replaced with the Ad2 hexon gene ORF in such a fashion so as to maintain the proper Ad5 sequences upstream and downstream of the hexon gene. Adenoviral vectors comprising modified or chimeric hexon proteins can be constructed by homologous recombination using standard techniques and human embryonic kidney 293 cells (see, e.g., Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra). For instance, map units 0 to 57.3 of dlAd5NCAT (Gall et al., supra) can be isolated by Bsu36I digestion, and map units 58.4 to 100 of dlAd5NCAT can be isolated by DrdI digestion. These DNA fragments can be transfected into 293 cells along with pH5-2.

A neutralizing antibody directed against the parental vector can be employed to facilitate the generation of hexon replacement constructs. For example, when replacing the loop 1 and loop 2 regions of an Ad5 vector with Ad7 loop sequences, anti-Ad5 neutralizing polyclonal or monoclonal antibodies (directed against the loops 1 and 2 of Ad5 hexon) can be added to a the medium of cells in which the chimeric vector is being propagated. presence of the Ad5 neutralizing antibodies substantially blocks the propagation of the undesired wildtype Ad5 vector(s), while the chimeric vector is unaffected. Furthermore, the recombinant vectors comprising a chimeric hexon ORF can be generated by homologous recombination using a plasmid that carries a marker gene, such as Green Fluorescent Protein (GFP), adjacent to the chimeric or novel hexon ORF (e.g., between the fiber and hexon genes). In this way, genomes that could harbor the chimeric hexon gene should also harbor the marker gene. The marker gene

would then be expressed as a late protein, so that cells that potentially comprise the desired adenoviral genome can be easily identified.

Similarly, vectors (particularly adenoviral vectors) can be constructed that have the aforementioned hexon modifications, and which have further modifications, for instance, in the adenoviral fiber coding sequences. This can be accomplished by making the hexon modifications described above, and using different parental plasmids for homologous recombination, such as parental plasmids comprising mutations in fiber coding sequences. In particular, the "5 base/7 fiber" vector can be employed as a starting vector for vector construction.

All of the viral vectors prepared according to this example can be plaque-purified, amplified, and further purified using standard methods (Rosenfeld et al. (1991), supra; Rosenfeld et al. (1992), supra).

#### Example 6

This example describes a characterization of the activity in vitro and in vivo of the vectors described in the preceding Examples.

Each of the viruses prepared as described in the preceding Examples can be evaluated in vitro and in vivo using standard methods as previously described (e.g., Kass-Eisler et al., <u>supra</u>), and as set forth herein. In particular, for the in vitro studies, the various vectors along with control vectors (e.g., the "pure 5" and "5 base/7 fiber" vectors, and the Ad5 wild-type vector) can be added to human lung carcinoma A549 cells alone, or in the presence of dilutions of serum from hosts infected with Ad5, Ad7, "pure 5" CAT vector, or "5 base/7 fiber" CAT vector, or anti-FLAG epitope serum. The cells are then evaluated for CAT activity to determine the ability

of antibodies present in the serum to block gene expression.

The in vivo studies can be carried out in Sprague-Dawley rats. The Sprague-Dawley rat as opposed to the mouse or cotton rat is preferred for these experiments since the rat is non-permissive, and the wild-type adenovirus cannot replicate in this host. Accordingly, immunizations can be carried out using wild-type viruses (e.g., wild-type Ad5 or Ad7), the "pure 5" CAT vector, and the "5 base/7 fiber" CAT vector by intravenous administration (e.g., Kass-Eisler et al., supra). At various times ranging from about one to about four weeks later, the vector of interest can be administered intravenously or directly into the airways of the host. Whereas intravenous administration allows an assessment of the "worst case scenario" (i.e., wherein the vector is in immediate contact with the circulating humoral immune system, and thus the strongest immune response is to be expected), introduction in the airways of the host allows an evaluation of a compartmentalized and mucosal humoral immune response.

CAT activity can be quantified as previously described in all the relevant organs, e.g., liver, heart, and lung for intravenous administration, and lung only for respiratory administration. Appropriate standards can be used to compensate for variations in organ expression of CAT activity (see e.g., Kass-Eisler et al., Gene Therapy, 2 395-402 (1994)). The in vitro and in vivo results can be compared and assessed using standard statistical methods.

All of the references cited herein, including the GenBank® Data Bank sequence information, are hereby incorporated in their entireties by reference.

WO 98/40509 PCT/US98/05033

57

While this invention has been described with emphasis upon preferred embodiments, it will be obvious to those of ordinary skill in the art that the preferred embodiments can be varied. It is intended that the invention can be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the appended claims.

58

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- (ii) TITLE OF INVENTION: CHIMERIC ADENOVIRAL COAT PROTEIN AND METHODS OF USING SAME
- (iii) NUMBER OF SEQUENCES: 56
- (iv) COMPUTER READABLE FORM:

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WO 98/40509

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		(E	3) CC	MPUI ERAI	ER:	IBM SYST	PC c EM: entIn	Ompa PC-E	tibl OS/M	IS-DC		ersi	.on #	1.30	(EPO	. · )	
	(vi)	(A	A) AF	PLIC	CATIC	N NU	ATA: IMBER 3-MA	t: US		1634	6		•				
(2)	INE	ORMA	MOITA	FOF	SEC	] ID	NO:1	. <b>:</b>	•								
	(i)	(A (E	() LE () TY () SI	NGTH PE: RAND	l: 29 nucl EDNE	07 b .eic	STIC ase acid doub	pair 1	s <sub>.</sub>			•					
	(ii)	MOL	ECUI	E TY	PE:	DNA	(gen	omic	2)					:			٠٠.
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							•										
	Ala									Ser				ATC Ile 15	Ser		48
														TTT Phe			96
														AAC Asn			144
														CGT Arg			192
														TCG Ser			240
														GAT Asp 95			288
														CCT Pro		•	336
														AAG Lys			384
														CGG Arg			432
														GAA Glu		٠	480

GAA GAG CAA AAC GCT CGA GAT CAG GCT ACT AAG AAA ACA CAT GTC TAT

170

Glu Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr

528

175

GCC Ala	CAG Gln	GCT Ala	CCT Pro 180	TTG Leu	TCT Ser	GGA Gly	GAA Glu	ACA Thr 185	ATT Ile	ACA Thr	AAA Lys	AGC Ser	GGG Gly 190	CTA Leu	CAA Gln	576
ATA Ile	GGA Gly	TCA Ser 195	GAC Asp	AAT Asn	GCA Ala	GAA Glu	ACA Thr 200	CAA Gln	GCT Ala	AAA Lys	CCT	GTA Val 205	TAC Tyr	GCA Ala	GAT Asp	624
CCT Pro	TCC Ser 210	Tyr	CAA Gln	CCA Pro	GAA Glu	CCT Pro 215	CAA Gln	ATT Ile	GGC Gly	GAA Glu	TCT Ser 220	CAG Gln	TGG Trp	AAC Asn	GAA Glu	672
GCT Ala 225	Asp	GCT Ala	AAT Asn	GCG Ala	GCA Ala 230	GGA Gly	GGG Gly	AGA Arg	GTG Val	CTT Leu 235	AAA Lys	AAA Lys	ACA Thr	ACT Thr	CCC Pro 240	720
ATG Met	AAA Lys	CCA Pro	TGC Cys	TAT Tyr 245	GGA Gly	TCT Ser	TAT Tyr	GCC Ala	AGG Arg 250	CCT Pro	ACA Thr	AAT Asn	CCT Pro	TTT Phe 255	GGT Gly	768
GGT Gly	CAA Gln	TCC Ser	GTT Val 260	CTG Leu	GTT Val	CCG Pro	GAT Asp	GAA Glu 265	AAA Lys	GGG Gly	GTG Val	CCT Pro	CTT Leu 270	CCA Pro	AAG Lys	816
GTT Val	GAC Asp	TTG Leu 275	CAA Gln	TTC Phe	TTC Phe	TCA Ser	AAT Asn 280	ACT Thr	ACC Thr	TCT Ser	TTG Leu	AAC Asn 285	GAC Asp	CGG Arg	CAA Gln	864
GGC Gly	AAT Asn 290	GCT Ala	ACT Thr	AAA Lys	CCA Pro	AAA Lys 295	GTG Val	GTT Val	TTG Leu	TAC Tyr	AGT Ser 300	GAA Glu	GAT Asp	GTA Val	AAT Asn	912
ATG Met 305	GAA Glu	ACC Thr	CCA Pro	GAC Asp	ACA Thr 310	CAT His	CTG Leu	TCT Ser	TAC Tyr	AAA Lys 315	CCT Pro	GGA Gly	AAA Lys	GGT Gly	GAT Asp 320	960
GAA Glu	AAT Asn	TCT Ser	AAA Lys	GCT Ala 325	ATG Met	TTG Leu	GGT Gly	CAA Gln	CAA Gln 330	TCT Ser	ATG Met	CCA Pro	AAC Asn	AGA Arg 335	CCC Pro	1008
AAT Asn	TAC Tyr	ATT Ile	GCT Ala 340	TTC Phe	AGG Arg	GAC Asp	AAT Asn	TTT Phe 345	ATT Ile	GGC Gly	CTA Leu	ATG Met	TAT Tyr 350	TAT Tyr	AAC Asn	1056
AGC Ser	ACT Thr	GGC Gly 355	AAC Asn	ATG Met	GGT Gly	GTT Val	CTT Leu 360	GCT Ala	GGT Gly	CAG Gln	GCA Ala	TCG Ser 365	CAG Gln	CTA Leu	AAT Asn	1104
GCC Ala	GTG Val 370	GTA Val	GAT Asp	TTG Leu	CAA Gln	GAC Asp 375	AGA Arg	AAC Asn	ACA Thr	GAG Glu	CTG Leu 380	TCC Ser	TAT Tyr	CAA Gln	CTC Leu	1152
TTG Leu 385	CTT Leu	GAT Asp	TCC Ser	ATA Ile	GGT Gly 390	GAT Asp	AGA Arg	ACC Thr	AGA Arg	TAT Tyr 395	TTT Phe	TCT Ser	ATG Met	TGG Trp	AAT Asn 400	1200
CAG Gln	GCT Ala	GTA Val	GAC Asp	AGC Ser 405	TAT Tyr	GAT Asp	CCA Pro	GAT Asp	GTT Val 410	AGA Arg	ATC Ile	ATT Ile	GAA Glu	AAC Asn 415	CAT His	1248
GGA Gly	ACT Thr	GAG Glu	GAT Asp 420	GAA Glu	TTG Leu	CCA Pro	AAT Asn	TAT Tyr 425	TGT Cys	TTT Phe	CCT Pro	CTT Leu	GGG Gly 430	GGT Gly	ATT Ile	1296

						-				
		Asp			AAG Lys	Asn			TCA Ser	1344
Gly					AAA Lys				ACA Thr	1392
					TTT Phe					1440
					TAC Tyr 490					1488
					ACC Thr					1536
					AAG Lys					1584
					GCG Ala		Ser			1632
					CAC His					1680
					CGC Arg 570				ATT Ile	1728
	${\tt Pro}$				AAA Lys					1776
					AGG Arg					1824
					AGA Arg					1872
					ACC Thr					1920
					CTC Leu 650				•	1968
					GCC Ala				ATA Ile	2016
					TCC Ser				TGG Trp	2064

GCA Ala	GCA Ala 690	TTT Phe	CGC Arg	GGT Gly	TGG Trp	GCC Ala 695	TTC Phe	ACA Thr	CGC Arg	TTG Leu	AAG Lys 700	ACA Thr	AAG Lys	GAA Glu	ACC Thr	2112
CCT Pro 705	TCC Ser	CTG Leu	GGA Gly	TCA Ser	GGC Gly 710	TAC Tyr	GAC Asp	CCT Pro	TAC Tyr	TAC Tyr 715	ACC Thr	TAC Tyr	TCT Ser	GGC Gly	TCC Ser 720	2160
ATA Ile	CCA Pro	TAC Tyr	CTT Leu	GAC Asp 725	GGA Gly	ACC Thr	TTC Phe	TAT Tyr	CTT Leu 730	AAT Asn	CAC His	ACC Thr	TTT Phe	AAG Lys 735	AAG Lys	2208
GTG Val	GCC Ala	ATT Ile	ACC Thr 740	TTT Phe	GAC Asp	TCT Ser	TCT Ser	GTT Val 745	AGC Ser	TGG Trp	CCG Pro	GGC Gly	AAC Asn 750	GAC Asp	CGC Arg	2256
CTG Leu	CTT Leu	ACT Thr 755	CCC Pro	AAT Asn	GAG Glu	TTT Phe	GAG Glu 760	ATT Ile	AAA Lys	CGC Arg	TCA Ser	GTT Val 765	GAC Asp	GGG Gly	GAG Glu	2304
GGC Gly	TAC Tyr 770	AAC Asn	GTA Val	GCT Ala	CAG Gln	TGC Cys 775	AAC Asn	ATG Met	ACC Thr	AAG Lys	GAC Asp 780	TGG Trp	TTC Phe	CTG Leu	GTG Val	2352
CAG Gln 785	ATG Met	TTG Leu	GCC Ala	AAC Asn	TAC Tyr 790	AAT Asn	ATT Ile	GGC Gly	TAC Tyr	CAG Gln 795	GGC Gly	TTC Phe	TAC Tyr	ATT Ile	CCA Pro 800	2400
GAA Glu	AGC Ser	TAC Tyr	AAG Lys	GAC Asp 805	CGC Arg	ATG Met	TAC Tyr	TCG Ser	TTC Phe 810	TTC Phe	AGA Arg	AAC Asn	TTC Phe	CAG Gln 815	CCC Pro	2448
ATG Met	AGC Ser	CGG Arg	CAA Gln 820	GTG Val	GTT Val	GAC . Asp	GAT Asp	ACT Thr 825	AAA Lys	TAC Tyr	AAG Lys	GAG Glu	TAT Tyr 830	CAG Gln	CAG Gln	2496
GTT Val	GGA Gly	ATT Ile 835	CTT	CAC His	CAG Gln	CAT His	AAC Asn 840	AAC Asn	TCA Ser	GGA Gly	TTC Phe	GTA Val 845	GGC Gly	TAC Tyr	CTC Leu	2544
GCT Ala	CCC Pro 850	ACC Thr	ATG Met	CGC Arg	GAG Glu	GGA Gly 855	CAG Gln	GCT Ala	TAC Tyr	CCC Pro	GCC Ala 860	AAC Asn	GTG Val	CCC Pro	TAC Tyr	2592
CCA Pro 865	CTA Leu	ATA Ile	GGC - Gly	AAA Lys	ACC Thr 870	GCG Ala	GTT Val	GAC Asp	AGT Ser	ATT Ile 875	ACC Thr	CAG Gln	AAA Lys	AAG Lys	TTT Phe 880	2640
CTT Leu	TGC Cys	GAT Asp	CGC Arg	ACC Thr 885	CTT Leu	TGG Trp	CGC Arg	ATC Ile	CCA Pro 890	TTC Phe	TCC Ser	AGT Ser	AAC Asn	TTT Phe 895	ATG Met	2688
TCC Ser	ATG Met	GGC Gly	GCA Ala 900	CTC Leu	ACA Thr	GAC Asp	CTG Leu	GGC Gly 905	CAA Gln	AAC Asn	CTT Leu	CTC Leu	TAC Tyr 910	GCC Ala	AAC Asn	2736
TCC Ser	GCC Ala	CAC His 915	GCG Ala	CTA Leu	GAC Asp	ATG Met	ACT Thr 920	TTT Phe	GAG Glu	GTG Val	GAT Asp	CCC Pro 925	ATG Met	GAC Asp	GAG Glu	2784
CCC Pro	ACC Thr 930	CTT Leu	CTT Leu	TAT Tyr	GTT Val	TTG Leu 935	TTT Phe	GAA Glu	GTC Val	TTT Phe	GAC Asp 940	GTG Val	GTC Val	CGT Arg	GTG Val	2832

CAC CAG CCG CAC CGC GGC GTC ATC GAG ACC GTG TAC CTG CGC ACG CCC 2880 His Gln Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro 960 950 955 2907 TTC TCG GCC GGC AAC GCC ACA ACA TAA Phe Ser Ala Gly Asn Ala Thr Thr 965 (2) INFORMATION FOR SEQ ID NO:2: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 968 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 100 Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly 120 Ala Pro Asn Ser Cys Glu Trp Glu Gln Thr Glu Asp Ser Gly Arg Ala Val Ala Glu Asp Glu Glu Glu Glu Asp Glu Glu Glu Glu Glu Glu 145 Glu Glu Gln Asn Ala Arg Asp Gln Ala Thr Lys Lys Thr His Val Tyr Ala Gln Ala Pro Leu Ser Gly Glu Thr Ile Thr Lys Ser Gly Leu Gln Ile Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp Pro Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu . 215

Ala Asp Ala Asn Ala Ala Gly Gly Arg Val Leu Lys Lys Thr Thr Pro

Met	Lys	Pro	Суѕ	Tyr 245	Gly	Ser	Tyr	Ala	Arg 250	Pro	Thr	Asn	Pro	Phe 255	Gly
Gly	Gln	Ser	Val 260	Leu	Val	Pro	Asp	Glu 265	Lys	Gly	Val	Pro	Leu 270	Pro	Lys
Val	Asp	Leu 275	Gln	Phe	Phe	Ser	Asn 280	Thr	Thr	Ser	Leu	Asn 285	Asp	Arg	Glr
Gly	Asn 290	Ala	Thr	Lys	Pro	Lys 295	Val	Val	Leu	Tyr	Ser 300	Glu	Asp	Val	Asr
Met 305	Glu	Thr	Pro	Asp	Thr 310	His	Leu	Ser	Tyr	Lys 315	Pro	Gly	Lys	Gly	Asp 320
Glu	Asn	Ser	Lys	Ala 325	Met	Leu	Gly	Gln	Gln 330	Ser	Met	Pro	Asn	Arg 335	Pro
Asn	Tyr	Ile	Ala 340	Phe	Arg	Asp	Asn	Phe 345	Ile	Gly	Leu	Met	Tyr 350	Tyr	Asn
Ser	Thr	Gly 355	Asn	Met	Gly	Val	Leu 360	Ala	Gly	Gln	Ala	Ser 365	Gln	Leu	Asn
Ala	Val 370	Val	Asp	Leu	Gln	Asp 375	Arg	Asn	Thr	Glu	Leu 380	Ser	Tyr	Gln	Leu
Leu 385	Leu	Asp	Ser	Ile	Gly 390	Asp	Arg	Thr	Arg	Tyr 395	Phe	Ser	Met	Trp	Asn 400
Gln	Ala	Val	Asp	Ser 405	Tyr	Asp	Pro	Asp	Val 410	Arg	Ile	Ile	Glu	Asn 415	His
Gly	Thr	Glu	Asp 420	Glu	Leu	Pro	Asn	Tyr 425	Суѕ	Phe	Pro	Leu	Gly 430	Gly	Ile
Gly	Val	Thr 435	Asp	Thr	Tyr	Gln	Ala 440	Ile	Lys	Ala	Asn	Gly 445	Asn	Gly	Ser
Gly	Asp 450	Asn	Gly	Asp	Thr	Thr 455	Trp	Thr	Lys	Asp	Glu 460	Thr	Phe	Ala	Thr
Arg 465	Asn	Glu	Ile	Gly	Val 470	Gly	Asn	Asn	Phe	Ala 475	Met	Glu	Ile	Asn	Leu 480
Asn	Ala	Asn	Leu	Trp 485	Arg	Asn	Phe	Leu	Tyr 490	Ser	Asn	Ile	Ala	Leu 495	Tyr
Leu	Pro	Asp	Lys 500	Leu	Lys	Tyr	Asn	Pro 505	Thr	Asn	Val	Glu	Ile 510	Ser	Asp
Asn	Pro	Asn 515	Thr	Tyr	Asp	Tyr	Met 520	Asn	Lys	Arg	Val	Val 525	Ala	Pro	Gly
Leu	Val 530	Asp	Cys	Tyr	Ile	Asn 535	Leu	Gly	Ala	Arg	Trp 540	Ser	Leu	Asp	Tyr
Met 545	Asp	Asn	Val	Asn	Pro 550	Phe	Asn	His	His	Arg 555	Asn	Ala	Gly	Leu	Arg 560
Tyr	Arg	Ser	Met	Leu 565	Leu	Gly	Asn	Gly	Arg 570	Tyr	Val	Pro	Phe	His 575	Ile

	•								0.5						
Gln	Val	Pro	Gln 580	Lys	Phe	Phe	Ala	Ile 585	Lys	Asn	Leu	Leu	Leu 590	Leu	Pro
Gly	Ser	Tyr 595	Thr	Tyr	Glu	Trp	Asn 600	Phe	Arg	Lys	Asp	Val 605	Asn	Met	Val
Leu	Gln 610	Ser	Ser	Leu	Gly	Asn 615	Asp	Leu	Arg	Val	Asp 620	Gly	Ala	Ser	Ile
Lys 625	Phe	Asp	Ser	Ile	Cys 630	Leu	Tyr	Ala	Thr	Phe 635	Phe	Pro	Met	Ala	His 640
Asn	Thr	Ala	Ser	Thr 645	Leu	Glu	Ala	Met	Leu 650	Arg	Asn	Asp	Thr	Asn 655	Asp
Gln	Ser	Phe ·	Asn 660	Asp	Tyr	Leu	Ser	Ala 665	Ala	Asn	Met	Leu	Tyr 670	Pro	Ile
Pro	Ala	Asn 675	Ala	Thr	Asn	Val	Pro 680	Ilė	Ser	Ile	Pro	Ser 685	Arg	Asn	Trp
Ala	Ala 690	Phe	Arg	Gly	Tṛp	Ala 695	Phe	Thr	Arg	Leu	Lys 700	Thr	Lys	Glu	Thr
Pro 705	Ser	Leu	Gly	Ser	Gly 710	Tyr	Asp	Pro	Tyr	Tyr 715	Thr	Tyr	Ser	Gly	Ser 720
Ile	Pro	Tyr	Leu	Asp 725	Gly	Thr	Phe	Tyr	Leu 730	Asn	His	Thr	Phe	Lys 735	Lys
Val	Ala	Ile	Thr 740	Phe	Asp	Ser	Ser	Val 745	Ser	Trp	Pro	Gly	Asn 750	Asp	Arg
Leu	Leu	Thr 755	Pro	Asn	Glu	Phe	Glu 760	Ile	Lys	Arg	Ser	Val 765	Asp	Gly	Glu
Gly	Tyr. 770	Asn	Val	Ala	Gln	Cys 775	Asn	Met	Thr	Lys	Asp 780	Trp	Phe	Leu	Val
Gln 785	Met	Leu	Ala	Asn	Tyr 790	Asn	Ile	Gly	Tyr	Gln 795	Gly	Phe	Tyr	lle	Pro 800
				805	ė				810					Gln 815	
Met	Ser	Arg	Gln 820	Val	Val	Asp	Asp	Thr 825	Lys	Tyr	Lys	Glu	Tyr 830	Gln	Gln
Val	Gly	11e 835	Leu	His	Gln	His	Asn 840	Asn	Ser	Gly	Phe	Val 845	Gly	Tyr	Leu
Ala	Pro 850	Thr	Met	Arg	Glu	Gly 855	Gln	Ala	Tyr	Pro	Ala 860		Val	Pro	Tyr
Pro 865	Leu	Ile	Gly	Lys	Thr 870	Ala	Val	Asp	Ser	Ile 875	Thr	Gln	Lys	Lys	Phe 880
Leu	Cys	Asp	Arg	Thr 885	Leu	Trp	Arg	Ile	Pro 890		Ser	Ser	Asn	Phe 895	Met
Ser	Met	Gly	Ala 900	Leu	Thr	Asp	Leu	Gly 905		Asn	Leu	Leu	Tyr 910		Asn

									66							
Ser	Ala	His 915	Ala	Leu	Asp	Met	Thr 920	Phe	Glu	Val	Asp	Pro 925	Met	Asp	Glu	
Pro	Thr 930	Leu	Leu	Tyr	Val	Leu 935	Phe	Glu	Val	Phe	Asp 940	Val	Val	Arg	Val	•
945					Gly 950 Ala			Glu	Thr	Val 955	Tyr	Leu	Arg	Thr	Pro 960	
(2)	INFO	ORMA!	rion	FOR	SEQ	ID N	NO:3:	:								
	(i)	( <i>I</i> (I	A) LE B) TY C) ST	ENGTI (PE: (RANI	HARAC H: 28 nucl DEDNI DGY:	358 k Leic ESS:	ase acio doub	pai:	rs							
	(ii)	MOI	LECUI	E T	PE:	DNA	(ger	nomi	=)							
Thr'		( Z ( E	3) LC	AME/F	ON:	951,	952	ature 2 : /no		'Xaa	can	be e	eithe	er Gl	ln, His	s, or
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTIC	on: s	SEQ I	D NO	3:						
ATG Met 1	GCT Ala	ACC Thr	CCT Pro	TCG Ser 5	ATG Met	ATG Met	CCG Pro	CAG Gln	TGG Trp 10	TCT Ser	TAC Tyr	ATG Met	CAC His	ATC Ile 15	TCG Ser	48
GGC Gly	CAG Gln	GAC Asp	GCC Ala 20	TCG Ser	GAG Glu	TAC Tyr	CTG Leu	AGC Ser 25	CCC Pro	GGG Gly	CTG Leu	GTG Val	CAG Gln 30	TTT Phe	GCC Ala	96
CGC Arg	GCC Ala	ACC Thr 35	GAG Glu	ACG Thr	TAC Tyr	TTC Phe	AGC Ser 40	CTG Leu	AAT Asn	AAC Asn	AAG Lys	TTT Phe 45	AGA Arg	AAC Asn	CCC Pro	144
ACG Thr	GTG Val 50	GCG Ala	CCT Pro	ACG Thr	CAC His	GAC Asp 55	GTG Val	ACC Thr	ACA Thr	GAC Asp	CGG Arg 60	TCC Ser	CAG Gln	CGT Arg	TTG Leu	192
								CGT Arg								. 240
AAG Lys	GCG Ala	CGG Arg	TTC Phe	ACC Thr 85	CTA Leu	GCT Ala	GTG Val	GGT Gly	GAT Asp 90	Asn	CGT Arg	GTG Val	CTG Leu	GAC Asp 95	ATG Met	288
GCT Ala	TCC Ser	ACG Thr	TAC Tyr 100	Phe	GAC Asp	ATC Ile	CGC Arg	GGC Gly 105	GTG Val	CTG Leu	GAC Asp	AGG Arg	GGC Gly 110	CCT Pro	ACT Thr	336
TTT Phe	AAG Lys	CCC Pro 115	TAC Tyr	TCT Ser	GGC Gly	ACT Thr	GCC Ala 120	TAC Tyr	AAC Asn	GCC Ala	CTG Leu	GCT Ala 125	CCC Pro	AAG Lys	GGT Gly	384
GCC Ala	CCA Pro 130	AAT Asn	CCT Pro	TGC Cys	GAA Glu	TGG Trp 135	GAT Asp	GAA Glu	GCT Ala	GCT Ala	ACT Thr 140	GCT Ala	CTT Leu	GAA Glu	ATA Ile	432

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AAC Asn 145	CTA Leu	GAA Glu	GAA Glu	GAG Glu	GAC Asp 150	Asp	GAC Asp	AAC Asn	GAA Glu	GAC Asp 155	GAA Glu	GTA Val	GAC Asp	GAG Glu	CAA Gln 160		480
							GTA Val										528
							ATT Ile										576
							TTT Phe 200										624
							ATT										672
							CCA Pro										720
							GGC Gly										768
							ATG Met										816
			Asn				TTG Leu 280									•	864
GAA Glu	GAT Asp 290	GTA Val	GAT Asp	ATA Ile	GAA Glu	ACC Thr 295	CCA Pro	GAC Asp	ACT Thr	CAT His	ATT Ile 300	TCT Ser	TAC Tyr	ATG Met	CCC		912
							CGA Arg										960
							GCT Ala										1008
							AAT Asn							Gln			1056
							GAT Asp 360						Thr				1104
		Gln					TCC Ser										1152
							GAC Asp								ATT Ile 400		1200

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ATT Ile	GAA Glu	AAT Asn	CAT His	GGA Gly 405	ACT Thr	GAA Glu	GAT Asp	GAA Glu	CTT Leu 410	CCA Pro	AAT Asn	TAC Tyr	TGC Cys	TTT Phe 415	CCA Pro		1248
CTG Leu	GGA Gly	GGT Gly	GTG Val 420	ATT Ile	AAT Asn	ACA Thr	GAG Glu	ACT Thr 425	CTT Leu	ACC Thr	AAG Lys	GTA Val	AAA Lys 430	CCT Pro	AAA Lys		1296
ACA Thr	GGT Gly	CAG Gln 435	GAA Glu	AAT Asn	GGA Gly	TGG Trp	GAA Glu 440	AAA Lys	GAT Asp	GCT Ala	ACA Thr	GAA Glu 445	TTT Phe	TCA Ser	GAT Asp		1344
AAA Lys	AAT Asn 450	GAA Glu	ATA Ile	AGA Arg	GTT Val	GGA Gly 455	AAT Asn	AAT Asn	TTT Phe	GCC Ala	ATG Met 460	GAA Glu	ATC Ile	AAT Asn	CTA Leu		1392
AAT Asn 465	GCC Ala	AAC Asn	CTG Leu	TGG Trp	AGA Arg 470	AAT Asn	TTC Phe	CTG Leu	TAC Tyr	TCC Ser 475	AAC Asn	ATA Ile	GCG Ala	CTG Leu	TAT Tyr 480		1440
TTG Leu	CCC Pro	GAC Asp	AAG Lys	CTA Leu 485	AAG Lys	TAC Tyr	AGT Ser	CCT Pro	TCC Ser 490	AAC Asn	GTA Val	AAA Lys	ATT Ile	TCT Ser 495	GAT Asp		1488
AAC Asn	CCA Pro	AAC Asn	ACC Thr 500	TAC Tyr	GAC Asp	TAC Tyr	ATG Met	AAC Asn 505	AAG Lys	CGA Arg	GTG Val	GTG Val	GCT Ala 510	CCC Pro	GGG Gly		1536
TTA Leu	GTG Val	GAC Asp 515	TGC Cys	TAC Tyr	ATT Ile	AAC Asn	CTT Leu 520	GGA Gly	GCA Ala	CGC Arg	TGG Trp	TCC Ser 525	CTT Leu	GAC Asp	TAT Tyr		1584
Met	GAC Asp 530	AAC Asn	GTC Val	AAC Asn	CCA Pro	TTT Phe 535	AAC Asn	CAC His	CAC His	CGC Arg	AAT Asn 540	GCT Ala	GGC Gly	CTG Leu	CGC Arg	*	1632
TAC Tyr 545	CGC Arg	TCA Ser	ATG Met	TTG Leu	CTG Leu 550	GGC Gly	AAT Asn	GGT Gly	CGC Arg	TAT Tyr 555	GTG Val	CCC Pro	TTC Phe	CAC His	ATC Ile 560		1680
CAG Gln	GTG Val	CCT Pro	CAG Gln	AAG Lys 565	TTC Phe	TTT Phe	GCC Ala	ATT Ile	AAA Lys 570	AAC Asn	CTC Leu	CTT Leu	CTC Leu	CTG Leu 575	CCG Pro		1728
GGC Gly	TCA Ser	TAC Tyr	ACC Thr 580	TAC Tyr	GAG Glu	TGG Trp	AAC Asn	TTC Phe 585	AGG Arg	AAG Lys	GAT Asp	GTT Val	AAC Asn 590	ATG Met	GTT Val		1776
CTG Leu	CAG Gln	AGC Ser 595	TCC Ser	CTA Leu	GGA Gly	AAT Asn	GAC Asp 600	CTA Leu	AGG Arg	GTT Val	GAC Asp	GGA Gly 605	GCC Ala	AGC Ser	ATT Ile		1824
AAG Lys	TTT Phe 610	GAT Asp	AGC Ser	ATT Ile	TGC Cys	CTT Leu 615	TAC Tyr	GCC Ala	ACC Thr	TTC Phe	TTC Phe 620	CCC Pro	ATG Met	GCC Ala	CAC His		1872
AAC Asn 625	ACC Thr	GCC Ala	TCC Ser	ACG Thr	CTT Leu 630	GAG Glu	GCC Ala	ATG Met	CTT Leu	AGA Arg 635	AAC Asn	GAC Asp	ACC Thr	AAC Asn	GAC Asp 640		1920
CAG Gln	TCC Ser	TTT Phe	AAC Asn	GAC Asp 645	TAT Tyr	CTC Leu	TCC Ser	GCC Ala	GCC Ala 650	AAC Asn	ATG Met	CTC Leu	TAC Tyr	CCT Pro 655	ATA Ile		1968

	•				*	פס	•					
			ACC Thr									2016
			GGC Gly									2064
			TCG Ser									2112
			GAT Asp						Lys			2160
			TTT Phe 725									2208
			AAC Asn									2256
			GCC Ala							GTA Val		2304
			AAC Asn							CCA Pro		2352
	Ser		GAC Asp							CCC Pro 800		2400
			GTĠ Val 805									2448
		Ile	CAC His									2496
			CGC Arg									2544
			AAG Lys									2592
			ACC Thr									2640
			CTC Leu 885									2688
			CTA Leu							GAG Glu	. •	2736

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CCC Pro	ACC Thr	CTT Leu 915	CTT Leu	TAT Tyr	GTT Val	TTG Leu	TTT Phe 920	GAA Glu	GTC Val	TTT Phe	GAC Asp	GTG Val 925	GTC Val	CGT Arg	GTG Val	2784
CAC His	CGG Arg 930	CCG Pro	CAC	CGC Arg	GGC Gly	GTC Val 935	ATC Ile	GAA Glu	ACC Thr	GTG Val	TAC Tyr 940	CTG Leu	CGC Arg	ACG Thr	CCC Pro	2832
						ннн Хаа		нн								2858

- (2) INFORMATION FOR SEQ ID NO:4:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 952 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (ix) FEATURE
    - (A) NAME/KEY: misc\_feature
      (B) LOCATION: 951,952
- (D) OTHER INFORMATION: /note= "Xaa can be either Gln, His, or Thr"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
- Met Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser
- Gly Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala
- Arg Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro
- Thr Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu
- Thr Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr
- Lys Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met
- Ala Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr 100
- Phe Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly
- Ala Pro Asn Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile
- Asn Leu Glu Glu Asp Asp Asp Asn Glu Asp Glu Val Asp Glu Gln
- Ala Glu Gln Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly
- Ile Asn Ile Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr

Pro	Lys	Tyr 195	Ala	Asp	Lys	Thr	Phe 200	Gln	Pro	Glu	Pro	Gln 205	Ile	Gly	Glu	
Ser	Gln 210	Trp	Tyr	Glu	Thr	Glu 215	Ile	Asn	His	Ala	Ala 220	Gly	Arg	Val	Leu	
Lys 225	Lys	Thr	Thr	Pro.	Met 230	Lys	Pro	Cys	Tyr	Gly 235	Ser	Tyr	Ala	Lys	Pro 240	
Thr	Asn	Glu	Asn	Gly 245	Gly	Gln	Gly	Ile	Leu 250	Val	Lys	Gln	Gln	Asn 255	Gly	
Lys	Leu	Glu	Ser 260	Gln	Val	Glu	Met	Gln 265	Phe	Phe	Ser	Thr	Thr 270	Glu	Ala	
Thr	Ala	Gly 275	Asn	Gly	Asp	Asn	Leu 280	Thr	Pro	Lys	Val	Val 285	Leu	Tyr	Ser	
Glu	Asp 290	Vaĺ	Asp	Ile	Glu	Thr 295	Pro	Asp	Thr	His	11e 300	Ser	Tyr	Met	Pro	
Thr 305	Ile	Lys	Glu	Gly	Asn 310	Ser	Arg	Glu	Leu	Met 315	Gly	Gln	Gln	Ser	Met 320	
Pro	Asn	Arg	Pro	Asn 325	Tyr	Ile	Ala	Phe	Arg 330	Asp	Asn	Phe	Ile	Gly 335		
Met	Tyr	Tyr	Asn 340	Ser	Thr	Gly	Asn	Met 345		Val	Leu	Ala	Gly 350	Gln	Ala	
Ser	Gln	Leu 355	Asn	Ala	Val	Val	Asp 360	Leu	Gln	Asp	Arg	Asn 365	Thr	Glu	Leu	
Ser	Tyr 370	Gln	Leu	Leu	Leu	Asp 375	Ser	Ile	Gly	Asp	Arg 380	Thr	Arg	Tyr	Phe	
Ser 385	Met	Trp	Asn	Gln	Ala 390	Val	Asp	Ser	Tyr	Asp 395	Pro	Asp	V <sub>a</sub> l	Arg	Ile 400	
Ile	Glu	Asn	His	Gly 405	Thr	Glu	Asp	Glu	Leu 410	Pro	Asn	Tyr	Cys	Phe 415		
Leu	Gly	Gly ·	Val 420	Ile	Asn	Thr	Glu	Thr 425	Leu	Thr	Lys	Val	Lys 430	Pro	Lys	
Thr	Gly	Gln 435	Glu	Asn	Gly	Trp	Glu 440	Lys	Asp	Ala	Thr	Glu 445	Phe	Ser	Asp	
Lys	Asn 450	.Glu	Ile	Arg	Val	Gly 455	Asn	Asn	Phe	Ala	Met 460	Glu	Ile	Asn	Leu	
Asn 465	Ala	Asn	Leu	Trp	Arg 470	Asn	Phe	Leu	Tyr	Ser 475	Asn	Ile	Ala	Leu	Tyr 480	
Leu	Pro	Asp	Lys	Leu 485	Lys	Tyr	Ser	Pro	Ser 490	Asn	Val	Lys	Ile	Ser 495	Asp	
Asn	Pro	Asn	Thr 500	Tyr	Asp	Tyr	Met	Asn 505	Lys	Arg	Val	Val	Ala 510	Pro	Gly	
Leu	Val	Asp 515	Cys	Tyr	Ile	Asn	Leu 520	Gly	Ala	Arg	Trp	Ser 525	Leu	Asp	Tyr	

Met	Asp 530	Asn	Val	. Asn	Pro	Phe 535	Asn	His	His	Arg	Asn 540	Ala	Gly	Leu	Arg	
Tyr 545	Arg	Ser	Met	Leu	Leu 550	Gly	Asn	Gly	Arg	Tyr 555		Pro	Phe	His	Ile 560	
Gln	Val	Pro	Gln	Lys 565	Phe	Phe	Ala	Ile	Lys 570	Asn	Leu	Leu	Leu	Leu 575	Pro	
Gly	Ser	Tyr	Thr 580	Tyr	Glu	Trp	Asn	Phe 585	Arg	Lys	Asp	Val	Asn 590	Met	Val	
Leu	Gln	Ser 595	Ser	Leu	Gly	Asņ	Asp 600	Leu	Arg	Val	Asp	Gly 605	Ala	Ser	Ile	
Lys	Phe 610	Asp	Ser	Ile	Cys	Leu 615	.Tyr	Ala	Thr	Phe	Phe 620	Pro	Met	Ala	His	
625					630				Leu	635					640	
				645					Ala 650					655		
			660					665	Ser				670		_	
		675					680		Arg			685				
	690					695			Tyr		700					
/05					710				Leu	715					720	
				725					Ser 730					735		
			740					745	Lys				750	-		
		755					760		Thr			765				
	110					775			Tyr		780			•		
85					790				Phe	795					800	
				805					Lys 810					815		
			820					825	Ser				830			
		835					840		Tyr			845				
,ro	Leu 850	Ile	Gly	Lys	Thr	Ala 855	Val	Asp	Ser	Ile	Thr 860	Gln ·	Lys	Lys	Phe	

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Leu 865	Cys	Asp	Arg	Thr	Leu 870	Trp	Arg	Ile	Pro	Phe 875	Ser	Ser	Asn	Phe	Met 880		
Ser	Met	Gly	Ala	Leu 885	Thr	Asp	Leu	Gly	Gln 890	Asn	Leu	·Leu	Tyr	Ala 895	Asn		
Ser	Ala	His	Ala 900	Leu	Asp	Met	Thr	Phe 905	Glu	Val	Asp	Pro	Met 910	Asp	Glu	٠	
Pro	Thr	Leu 915	Leu	Tyr	Val	Leu	Phe 920	Glu	Val	Phe	Asp	Val 925	Val	Arg	Val		. •
His	Arg 930	Pro	His	Arg	Gly	Val 935	Ile	Glu	Thr	Val	Tyr 940	Leu	Arg	Thr	Pro		
Phe 945	Ser	Ala	Gly	Asn	Ala 950	Xaa	Xaa ,	·						. ,	•		•
(2)	INFO	ORMA'	rion	FOR	SEQ	ID N	NO:5:	•						·:			
	(i)	(	A) LI B) T: C) S:	CE CI ENGTI YPE: TRANI DPOLO	nuc] DEDNI	)3 ba Leic ESS:	ase p acid doub	oairs 1	3								
	(ii)	MOI	LECUI	LE T	YPE:	DNA	(ger	nomi	<b>=)</b>								
	(xi)	SE	QUEN	CE DI	ESCR	[PTI	ON: S	SEQ :	ID NO	0:5:							
				GAA Glu 5													48
				GAA Glu											_		96
				CAG Gln													144
CCT Pro	TTG Leu 50	TCT Ser	GGA Gly	GAA Glu	ACA Thr	ATT Ile 55	ACA Thr	AAA Lys	AGC Ser	GGG Gly	CTA Leu 60	CAA Gln	ATA Ile	GGA Gly	TCA Ser		192
				ACA Thr													240
				CAA Gln 85						Trp					Ala		288
AAT Asn	GCG Ala	GCA Ala	GGA Gly 100	GGG	AGA Arg	GTG Val	CTT	AAA Lys 105	Lys	ACA Thr	ACT	CCC	ATG Met 110	Lys	CCA Pro		336
			Ser	TAT Tyr				Thr					Gly		TCC		384

GTT Val	CTG Leu 130	Val	CCG Pro	GAT Asp	GAA Glu	AAA Lys 135	GGG Gly	GTG Val	CCT Pro	CTT	CCA Pro 140	AAG Lys	GTT Val	GAC Asp	TTG Leu	432
CAA Gln 145	Phe	TTC Phe	TCA Ser	AAT Asn	ACT Thr 150	ACC Thr	TCT Ser	TTG Leu	AAC Asn	GAC Asp 155	CGG Arg	CAA Gln	GGC Gly	AAT Asn	GCT Ala 160	480
ACT Thr	AAA Lys	CCA Pro	AAA Lys	GTG Val 165	GTT Val	TTG Leu	TAC Tyr	AGT Ser	GAA Glu 170	GAT Asp	GTA Val	AAT Asn	ATG Met	GAA Glu 175	ACC Thr	528
CCA Pro	GAC Asp	ACA Thr	CAT His 180	CTG Leu	TCT Ser	TAC Tyr	AAA Lys	CCT Pro 185	GGA Gly	AAA Lys	GGT Gly	GAT Asp	GAA Glu 190	AAT Asn	TCT Ser	576
AAA Lys	GCT Ala	ATG Met 195	TTG Leu	GGT Gly	CAA Gln	CAA Gln	TCT Ser 200	ATG Met						•		603
(2')	INF	ORMA!	TION	FOR	SEQ	ID 1	10:6:	:								
	(i	() ()	A) LI B) T	ENGTI YPE:	H: 20	CTERI D1 am no ac line	nino cid		ds							
	(ii	) MOI	LECUI	LE T	YPE:	pept	ide									
	(xi	) SE(	QUENC	CE DI	ESCR	PTIC	on: s	SEQ ]	D NO	0:6:						
Ser 1	Cys	Glu	Trp	Glu 5	Gln	Thr	Glu	Asp	Ser 10	Gly	Arg	Ala	Val	Ala 15	Glu	
Asp	Glu	Glu	Glu 20	Glu	Asp	Glu	Asp	Glu 25	Glu	Glu	Glu	Glu	Glu 30	Glu	Gln	
Asn	Ala	Arg 35	Asp	Gln	Ala	Thr	Lys 40	Lys	Thr	His	Val	Tyr 45	Ala	Gln	Ala	
Pro	Leu 50	Ser	Gly	Glu	Thr	Ile 55	Thr	Lys	Ser	Gly	Leu 60	Gln	Ile	Gly	Ser	
Asp 65	Asn	Ala	Glu	Thr	Gln 70	Ala	Lys	Pro	Val	Tyr 75	Ala	Asp	Pro	Ser	Tyr 80	
Gln	Pro	Glu	Pro	Gln 85	Ile	Gly	Glu	Ser	Gln 90	Trp	Asn	Glu	Ala	Asp 95	Ala	
Asn	Ala	Ala	Gly 100	Gly	Arg	Val	Leu	Lys 105	Lys	Thr	Thr	Pro	Met 110	Lys	Pro	
Cys	Tyr	Gly 115	Ser	Tyr	Ala	Arg	Pro 120	Thr	Asn	Pro	Phe	Gly 125	Gly	Gln	Ser	
Val	Leu 130	Val	Pro	Asp	Glu	Lys 135	Gly	Val	Pro	Leu	Pro 140	Lys	Val	Asp	Leu	
Gln 145	Phe	Phe	Ser	Asn	Thr 150	Thr	Ser	Leu	Asn	Asp 155	Arg	Gln	Gly	Asn	Ala 160	
Thr	Lys	Pro	Lys	Val 165	Val	Leu	Tyr		Glu 170	Asp	Val	Asn	Met	Glu 175	Thr	

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Pro Asp Thr His Leu Ser Tyr Lys Pro Gly Lys Gly Asp Glu Asn Ser 185

Lys Ala Met Leu Gly Gln Gln Ser Met . 195 200

### (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
   (A) LENGTH: 567 base pairs
   (B) TYPE: nucleic acid

  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

				GAT Asp 5													48
				GAC Asp													96
				GTA Val													144
				ATT Ile													192
				TTT Phe													240
TAC Tyr	GAA Glu	ACT Thr	GAA Glu	ATT Ile 85	AAT Asn	CAT	GCA Ala	GCT Ala	GGG Gly 90	AGA Arg	GTC Val	CTT Leu	AAA Lys	AAG Lys 95	ACT Thr		288
				CCA Pro												-	336
				GGC Gly											GAA Glu		384
				ATG Met													432
				TTG Leu							Tyr				GTA Val 160	,	480
				CCA Pro 165													528
GAA	GGT	AAC	TCA	CGA	GAA	CTA	ATG	GGC	CAA	CAA	TCT	ATG	•		•		567

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180 185

- (2) INFORMATION FOR SEQ ID NO:8:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 189 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu 1 5 10.

Glu Glu Asp Asp Asp Asp Glu Asp Glu Val Asp Glu Gln Ala Glu Gln 20 . 25 30

Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile 35 40

Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr Pro Lys Tyr 50 55

Ala Asp Lys Thr Phe Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp 65 70 75 80

Tyr Glu Thr Glu Ile Asn His Ala Ala Gly Arg Val Leu Lys Lys Thr 85 90

Thr Pro Met Lys Pro Cys Tyr Gly Ser Tyr Ala Lys Pro Thr Asn Glu 100 105 110

Asn Gly Gly Gln Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu 115 120 125

Ser Gln Val Glu Met Gln Phe Phe Ser Thr Thr Glu Ala Thr Ala Gly 130 135 140

Asn Gly Asp Asn Leu Thr Pro Lys Val Val Leu Tyr Ser Glu Asp Val 145 150 155 160

Asp Ile Glu Thr Pro Asp Thr His Ile Ser Tyr Met Pro Thr Ile Lys 165 170 175

Glu Gly Asn Ser Arg Glu Leu Met Gly Gln Gln Ser Met 180 185

- (2) INFORMATION FOR SEQ ID NO:9:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 153 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ACC GAA GAT AGC GGC CGG GCA GTT GCC GAG GAT GAA GAA GAA GAT

Thr 1	Glu	Asp	Ser	Gly 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15	Asp		
								GAG Glu 25									96
								CAG Gln									144
	ACA Thr 50							٠		-).	·				.· ·		153
(2)	INFO	RMAT	NOI	FOR	SEQ	ID i	NO:10	): ·		,							•
	(i)	( <i>I</i>	A) LE 3) TY	CE CHENGTH (PE: (POLC	4: 51 am <b>i</b> r	l ami	ino a	CS: acids	3				,	:			
	(ii)	MOI	LECUI	E TY	PE:	pept	cide										
	(xi)	SEC	QUENC	CE DE	ESCR	PTI	ON: S	SEQ I	ID NO	0:10							
Thr 1	Glu	Asp	Ser	Gly. 5	Arg	Ala	Val	Ala	Glu 10	Asp	Glu	Glu	Glu	Glu 15			
Glu	Asp	Glu	Glu 20	Glu	Glu	Glu	Glu	Glu 25	Gln	· Asn	Ala	Arg	Asp 30	Gln	Ala		
Thr	_	Lys 35	Thr	His	Val	Tyr	Ala 40	Gln	Ala	Pro	Leu	Ser 45	Gly	Glu	Thr		•
Ile	Thr 50	Lys									٠.			• •		•	
(2)	INFO	ORMA	NOIT	FOR	SEQ	ID i	NO:1	1:									•
	(i)	() ()	A) LE B) TY C) ST	CE CI ENGTI YPE: TRANI DPOLO	nuc. DEDNI	35 baleic ESS:	ase pacion doub	pair: d	5			•					
	(ii	MOI	LECUI	LE T	YPE:	DNA	(ge	nomi	c)								
	(xi	) SE	QUENC	CE DI	ESCR:	IPTI	ON:	SEQ :	ID N	0:11	:						
								CTA Leu		Glu					AAC Asn		48
								GAG Glu 25						Val	TTT Phe		96
			Pro					AAT Asn									135

- (2) INFORMATION FOR SEQ ID NO:12:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 45 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ala Ala Thr Ala Leu Glu Ile Asn Leu Glu Glu Glu Asp Asp Asn 1 5 10 15

Glu Asp Glu Val Asp Glu Gln Ala Glu Gln Gln Lys Thr His Val Phe
20 25 30

Gly Gln Ala Pro Tyr Ser Gly Ile Asn Ile Thr Lys Glu 35 40 45

- (2) INFORMATION FOR SEQ ID NO:13:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 33 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCA GAC AAT GCA GAA ACA CAA GCT AAA CCT GTA Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val 1 5 10

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 11 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val 1 5 10

- (2) INFORMATION FOR SEQ ID NO:15:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 21 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

	79
	GAA GGT CAA ACA CCT AAA Glu Gly Gln Thr Pro Lys 5
(2)	INFORMATION FOR SEQ ID NO:16:
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 7 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
Val 1	Glu Gly Gln Thr Pro Lys 5
(2)	INFORMATION FOR SEQ ID NO:17:
	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 24 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: double</li> <li>(D) TOPOLOGY: linear</li> </ul>
	(ii) MOLECULE TYPE: DNA (genomic)
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
	GAA GCT GAT GCT AAT GCG GCA Glu Ala Asp Ala Asn Ala Ala 5
(2)	INFORMATION FOR SEQ ID NO:18:
	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 8 amino acids</li><li>(B) TYPE: amino acid</li><li>(D) TOPOLOGY: linear</li></ul>
	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
Asn 1	Glu Ala Asp Ala Asn Ala Ala 5
(2)	INFORMATION FOR SEQ ID NO:19:
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 24 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: DNA (genomic)
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:
	GAA ACT GAA ATT AAT CAT GCA Glu Thr Glu Ile Asn His Ala 5

24

42

- (2) INFORMATION FOR SEQ ID NO:20:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 8 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Tyr Glu Thr Glu Ile Asn His Ala

- (2) INFORMATION FOR SEQ ID NO:21:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 42 base pairs

    - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TCC GTT CTG GTT CCG GAT GAA AAA GGG GTG CCT CTT CCA AAG Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys

- (2) INFORMATION FOR SEQ ID NO:22:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 14 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ser Val Leu Val Pro Asp Glu Lys Gly Val Pro Leu Pro Lys 1 5

- (2) INFORMATION FOR SEQ ID NO:23:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 42 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GGC ATT CTT GTA AAG CAA CAA AAT GGA AAG CTA GAA AGT CAA Gly Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln

- (2) INFORMATION FOR SEQ ID NO:24:
  - (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 14 amino acids

	81
	(B) TYPE: amino acid (D) TOPOLOGY: linear
••	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
1 y	Ile Leu Val Lys Gln Gln Asn Gly Lys Leu Glu Ser Gln 5 10
2)	INFORMATION FOR SEQ ID NO:25:
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 51 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: DNA (genomic)
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
	AAT ACT ACC TCT TTG AAC GAC CGG CAA GGC AAT GCT ACT AAA CCA Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro 5 10 15
AA ys	5:
2)	INFORMATION FOR SEQ ID NO:26:
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 17 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: peptide
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:
er 1	Asn Thr Thr Ser Leu Asn Asp Arg Gln Gly Asn Ala Thr Lys Pro 5 10 15
ys	
2)	INFORMATION FOR SEQ ID NO:27:
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 48 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: linear
:	(ii) MOLECULE TYPE: DNA (genomic)
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
	ACT ACT GAG GCG ACC GCA GGC AAT GGT GAT AAC TTG ACT CCT AAA Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys 5 10 15

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 amino acids
- (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Ser Thr Thr Glu Ala Thr Ala Gly Asn Gly Asp Asn Leu Thr Pro Lys 1 5 10 15

- (2) INFORMATION FOR SEQ ID NO:29:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29: .

TTG TAC AGT GAA GAT GTA AAT ATG Leu Tyr Ser Glu Asp Val Asn Met

24

- (2) INFORMATION FOR SEQ ID NO:30:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 8 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ: ID NO:30:

Leu Tyr Ser Glu Asp Val Asn Met
1 5

- (2) INFORMATION FOR SEQ ID NO:31:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

TTG TAC AGT GAA GAT GTA GAT ATA Leu Tyr Ser Glu Asp Val Asp Ile 1 5

- (2) INFORMATION FOR SEQ ID NO:32:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 8 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear

PCT/US98/05033

- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Leu Tyr Ser Glu Asp Val Asp Ile 1 5

- (2) INFORMATION FOR SEQ ID NO:33:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 36 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGA AAA GGT GAT GAA AAT TCT AAA GCT ATG TTG GGT Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly

36

- (2) INFORMATION FOR SEQ ID NO:34:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 12 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
- Gly Lys Gly Asp Glu Asn Ser Lys Ala Met Leu Gly
  1 5 10
- (2) INFORMATION FOR SEQ ID NO:35:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 36 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ACT ATT AAG GAA GGT AAC TCA CGA GAA CTA ATG GGC
Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly
1 5 10

- (2) INFORMATION FOR SEQ ID NO:36:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 12 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Thr Ile Lys Glu Gly Asn Ser Arg Glu Leu Met Gly

- (2) INFORMATION FOR SEQ ID NO:37:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 165 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

AAT Asn 1	TAT Tyr	TGT Cys	TTT Phe	CCT Pro 5	CTT Leu	GGG Gly	GGT Gly	ATT Įle	GGG Gly 10	GTA Val	ACT Thr	GAC Asp	ACC Thr	TAT. Tyr 15	CAA Gln	48
GCT Ala	ATT Ile	AAG Lys	GCT Ala 20	AAT Asn	GGC Gly	AAT Asn	Gly	TCA Ser 25	GGC Gly	GAT Asp	AAT Asn	GGA Gly	GAT Asp 30	ACT Thr	ACA Thr	96
TGG Trp	ACA Thr	AAA Lys 35	GAT Asp	GAA Glu	ACT Thr	TTT Phe	GCA Ala 40	ACA Thr	CGT Arg	AAT Asn	GAA Glu	ATA Ile 45	GGA Gly	GTG Val	GGT Gly	144
					GAA Glu											165

- (2) INFORMATION FOR SEQ ID NO:38:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 55 amino acids(B) TYPE: amino acid

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Asn Tyr Cys Phe Pro Leu Gly Gly Ile Gly Val Thr Asp Thr Tyr Gln

Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly Asp Asn Gly Asp Thr Thr 20

Trp Thr Lys Asp Glu Thr Phe Ala Thr Arg Asn Glu Ile Gly Val Gly

Asn Asn Phe Ala Met Glu Ile

- (2) INFORMATION FOR SEQ ID NO:39:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 153 base pairs (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)

									85								
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTIC	on: s	SEQ I	D NO	39:	:						
	TAC Tyr				Leu												48
	GTA Val																96
	GAA Glu																144
	GAA Glu 50	Ile	•	٠				9 "	•								153
(2)	INFO	RMAT	CION	FOR	SEQ	ID i	10:40	): ·	•								
	(i)	( <i>I</i>	QUENC A) LE B) TY O) TO	ENGTI	l: 51 amir	l ami	ino a		<b>3</b>				٠				
	(ii)	MOI	LECUI	LE TY	PE:	pept	ide						•			•	
	(xi)	SEC	QUENC	CE DE	ESCRI	[PTIC	on: s	SEQ :	ID NO	0:40	:						
Asn 1	Tyr	Cys	Phe	Pro 5	Leu	Gly	Gly	Val	Ile 10	Asn	Thr	Glu	Thr	Leu 15	Thr		
Lys	Val	Lys	Pro 20	Lys	Thr	Gly	Gln	Glu 25	Asn	Gly	Trp	Glu	Lys 30	_	Ala		
Thr	Glu	Phe 35	Ser	Asp	Lys	Asn	Glu 40	Ile	Arg	Val	Gly	Asn 45	Asn	Phe	Ala		
Met	Glu 50	Ile				•	•		. ·								
(2)	INFO	RMA	rion	FOR	SEQ	ID 1	NO: 4	1:									
	(i)	( <i>I</i> (I	QUENCA) LE B) TY C) SY D) TO	engti (PE: [Rani	h: 54 nucl	4 bas leic ESS:	se pa acio doul	airs d									•
	(ii)	MOI	LECUI	LE T	PE:	DNA	(ge	nomi	c) -								
	(xi)	SEÇ	QUENC	CE DI	ESCR	IPTI	ON:	SEQ :	ID N	0:41	:						
	ACT Thr																48
	AAT Asn							•					•	•			54
(2)	TND		TT 0.11	T0.D	ano	<b>-</b>	NO . 4	٠.									

- (2) INFORMATION FOR SEQ ID NO:42:
  - (i) SEQUENCE CHARACTERISTICS:
    (A) LENGTH: 18 amino acids

- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser Gly

1 10 15

Asp Asn

- (2) INFORMATION FOR SEQ ID NO:43:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 87 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

AAT ACA GAG ACT CTT ACC AAG GTA AAA CCT AAA ACA GGT CAG GAA AAT
Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn
1 5 10

GGA TGG GAA AAA GAT GCT ACA GAA TTT TCA GAT AAA AAT
Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn
20
25

- (2) INFORMATION FOR SEQ ID NO:44:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 29 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Asn Thr Glu Thr Leu Thr Lys Val Lys Pro Lys Thr Gly Gln Glu Asn 1 5 10 15

Gly Trp Glu Lys Asp Ala Thr Glu Phe Ser Asp Lys Asn 20 .25

- (2) INFORMATION FOR SEQ ID NO:45:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 21 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

ACT TTT GCA ACA CGT AAT GAA Thr Phe Ala Thr Arg Asn Glu

21

- (2) INFORMATION FOR SEQ ID NO:46:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Thr Phe Ala Thr Arg Asn Glu 1

- (2) INFORMATION FOR SEQ ID NO:47:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

ACA GAA TTT TCA GAT AAA AAT GAA Thr Glu Phe Ser Asp Lys Asn Glu

24

- (2) INFORMATION FOR SEQ ID NO:48:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 8 amino acids(B) TYPE: amino acid

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

Thr Glu Phe Ser Asp Lys Asn Glu

- (2) INFORMATION FOR SEQ ID NO:49:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: other nucleic acid
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

GAC TAC AAA GAC GAC GAC AAA Asp Tyr Lys Asp Asp Asp Asp Lys

- (2) INFORMATION FOR SEQ ID NO:50:
  - (i) SEQUENCE CHARACTERISTICS:

		(	A) L: B) T D) T	YPE:	ami	no a		cids									
	(ii)	) MO:	LECU:	LE T	YPE:	pep	tide	.,									
	(xi)	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ :	ID N	0:50	:						
Asp 1	Tyr	Lys	Asp	Asp 5	Asp	Asp	Lys										
(2)	INFO	ORMA'	rion	FOR	SEQ	ID 1	NO:5	1:									
	(i)	() ()	A) Li B) T	ENGT: YPE: TRANI	H: 2: nuci DEDNI	907 1 leic ESS:	ISTICoase acid doub ear	pai: i	rs								
	(ii)	MO	LECUI	LE T	YPE:	DNA	(ger	nomi	<b>=</b> )	•							
	(xi)	SE	QUENC	CE DI	ESCR:	IPTI	: : NC	SEQ :	ID NO	0:51	:						
ATG	GCT Ala 1	ACC Thr	CCT Pro	TCG Ser	ATG Met 5	ATG Met	CCG Pro	CAG Gln	TGG Trp	TCT Ser 10	TAC Tyr	ATG Met	CAC His	ATC Ile	TCG Ser 15		8
GGC Gly	CAG Gln	GAC Asp	GCC Ala	TCG Ser 20	GAG Glu	TAC Tyr	CTG Leu	AGC Ser	CCC Pro 25	GGG Gly	CTG Leu	GTG Val	CAG Gln	TTT Phe 30	GCC Ala	Ş	6
CGC Arg	GCC Ala	ACC Thr	GAG Glu 35	ACG Thr	TAC Tyr	TTC Phe	AGC Ser	CTG Leu 40	AAT Asn	AAC Asn	AAG Lys	TTT Phe	AGA Arg 45	AAC Asn	CCC Pro	14	4
ACG Thr	GTG Val	GCA Ala 50	CCT Pro	ACG Thr	CAC His	GAC Asp	GTA Val 55	ACC Thr	ACA Thr	GAC Asp	CGG Arg	TCC Ser 60	CAG Gln	CGT Arg	TTG Leu	19	12
ACG Thr	CTG Leu 65	CGG Arg	TTC Phe	ATC Ile	CCT Pro	GTG Val 70	GAC Asp	CGC Arg	GAG Glu	GAT Asp	ACC Thr 75	GCG Ala	TAC Tyr	TCG Ser	TAC Tyr	24	0
AAA Lys 80	GCG Ala	CGG Arg	TTC Phe	ACC Thr	CTG Leu 85	GCT Ala	GTG Val	GGT Gly	GAC Asp	AAC Asn 90	CGT Arg	GTG Val	CTT Leu	GAT Asp	ATG Met 95	28	8
GCT Ala	TCC Ser	ACG Thr	TAC	TTT Phe 100	GAC Asp	ATC Ile	CGC Arg	GGC Gly	GTG Val 105	CTG Leu	GAC Asp	AGG Arg	GGG Gly	CCT Pro 110	ACT Thr	33	6
TTT Phe	AAG Lys	CCC Pro	TAC Tyr 115	TCC Ser	GGC Gly	ACT Thr	GCC Ala	TAC Tyr 120	AAC Asn	GCT Ala	CTA Leu	GCT Ala	CCC Pro 125	AAG Lys	GGC Gly	38	34
GCT Ala	CCT Pro	AAC Asn 130	TCC Ser	TGT Cys	GAG Glu	TGG Trp	GAA Glu 135	CAA Gln	ACC Thr	GAA Glu	GAT Asp	AGC Ser 140	GGC Gly	CGG Arg	GCA Ala	43	32
GTT Val	GCC Ala 145	GAG Glu	GAT Asp	GAA Glu	GAA Glu	GAG Glu 150	GAA Glu	GAT Asp	GAA Glu	GAT Asp	GAA Glu 155	GAA Glu	GAG Glu	GAA Glu	GAA Glu	4.8	30

GAG Glu									528
CAG Gln				Thr			Gly		576
GGA Gly									624
TCC Ser									672
GAT Asp 225									720
AAA Lys									768
CAA Gln									816
GAC Asp									864
AAT Asn	Thr								912
GAA Glu 305									960
AAT Asn									1008
TAC Tyr									1056
ACT Thr									1104
GTG Val									1152
CTT Leu 385									1200
						Ile		CAT His 415	1248

GLY Thr Glu Asp Glu Leu Pro Asn Tyr Cys Phe Pro Leu Gly Gly Ile 425  GGG GTA ACT GAC ACC TAT CAA GCT ATT AAG GCT AAT GGC AAT GGC TCA Gly Val Thr Asp Thr Tyr Gln Ala Ile Lys Ala Asn Gly Asn Gly Ser 445  GGC GAT AAT GGA GAT ACT ACA TGG ACA AAA GAT GAA ACT TTT GCA ACA Gly Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr 450  CGT AAT GAA ATA GGA GTG GGT AAC AAC ATT GCC ATG GAA ATT AAC CTA Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu 470  AAT GCC AAC CTA TGG AGA AAT TTC CTT TAC TCC AAT ATT GCC TG TAC ASN Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr 480  CTG CAG ACA CAC CTA AGA ATA TAC AC CCC ACC AAT GTG GAA ATT TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 500  AAC CCC AAC ACC TAC GAC TAC ATG AAC AAG CAC GAT GTG GAC TAC Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 515  CTT GTA GAC TGC TAC ATT AAC CTT GGG GCG CCC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CAC GAT GCG GCC CTC Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 545  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CCC ATT CCC CTC CT											-							
GLY Val Thr Asp Thr Tyr Gln Ala ILe Lys Ala Asn Gly Asn Gly Ser 435  GGC GAT AAT GGA GAT ACT ACA TGG ACA AAA GAT GAA ACT TTT GCA ACA GLY Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr 450  CGT AAT GAA ATA GGA GAT GGT GGT ACA ACA CTT GCC ATG GAA ATT AAC CTA ARG AAT GAS GLU ILe Asn Leu 475  CGT AAT GAA ATA GGA GTG GGT ACA AAT TTC CCT TAC TCC AAT ATT GCC CTG TAC ARG AAT ASN ALA ASN Leu Trp Arg Asn Fhe Leu Tyr Ser Asn Ile Ala Leu Tyr 480  AAT GCC AAC CTA TGG AGA AAT TTC CTT TAC TCC AAT ATT GCC CTG TAC ASN ALA ASN Leu Trp Arg Asn Fhe Leu Tyr Ser Asn Ile Ala Leu Tyr 480  CTG CCA GAC AAC CTA AAA TAC AAC CCC ACC AAT GTG GAA ATT TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 500  AAC CCC AAC ACC TAC GAC TAC ATG AAC AGC GCA GTG GTG GCT CCC GGG Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 515  CTT GTA GAC TGC TAC ATT AAC CTT GGG GCG CGC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CGC AAT GGG GCC CTC CGT 16  Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 555  TAT CGC TCC ATG TTG TTG GGA AAC GGC CGC TAC GTG CCC TCT CTG TYR ATG SER Met Leu Leu Leu Leu Leu Leu Leu Cy Asp Noval Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Leu Pro 580  GGC TCA TAT ACA TAT GAA TGG AAC TCT AGA GAT GAG GAT GTG TT CAC ATT TYR Arg Ser Met Leu Leu Gly Asn Phe Arg Lys Asp Val Asn Met Val 600  GGC TCA TAT ACA TAT GAA TGG AAC TCT AGA GAT GAG GAT GTT AAC ATG GTT TYR Arg Ser Met Leu Leu Gly Asn Ang Lys Asp Val Asn Met Val 600  GGC TCA TAT ACA TAT GAA TGG AAC TCT AGA GAT GAC GGG GCT AGC ATT TYR TRY Glu Trp Asn Phe Ala Ile Lys Asn Leu Leu Leu Leu Pro 580  GGC TCA TAT ACA TAT GAA TGG AAC GAC TCT AGA GAT GAC GGG GCT AGC ATT 18: 18: 18: 18: 18: 18: 18: 18: 18: 18:		GGA Gly	ACT Thr	GAG Glu	GAT Asp	Glu	TTG Leu	CCA Pro	AAT Asn	TAT Tyr	Cys	TTT Phe	CCT Pro	CTT Leu	GGG Gly	Gly	ATT Ile	1296
Gly Asp Asn Gly Asp Thr Thr Trp Thr Lys Asp Glu Thr Phe Ala Thr 450  CGT AAT GAA ATA GGA GTG GGT AAC AAC TTT GCC ATG GAA ATT AAC CTA Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu 470  AAT GCC AAC CTA TGG AGA AAT TTC CTT TAC TCC AAT ATT GGC CTG TAC Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr 480  CTG CCA GAC AAG CTA AAA TAC AAC CCC ACC AAT GTG GAA ATA TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 510  AAC CCC AAC ACC TAC GAC TAC ATG AAC AAC CCC ACC AAT GTG GAA ATA TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 511  AAC CCC AAC ACC TAC GAC TAC ATG AAC AAG CGA GTG GTG GTG GCT CCC GGG Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 515  CTT GTA GAC TGC TAC ATT AAC CTT GGG GCG CGC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CAC AAT GCG GGC CTC CGT Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 545  TAT CCC TCC ATG TTG TTG GGA AAC GCC CGC TAC GTG CCC TTT CAC ATT Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 560  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTC CTC CTC CTG GAC TAC TAT ACC CTC CTC CTC CTC CTC CTC CTC CTC CTC C		GGG Gly	GTA Val	ACT Thr	Asp	ACC Thr	TAT Tyr	CAA Gln	GCT Ala	Ile	AAG Lys	GCT Ala	AAT Asn	GGC Gly	Asn	GGC Gly	TCA Ser	1344
Arg Asn Glu Ile Gly Val Gly Asn Asn Phe Ala Met Glu Ile Asn Leu 475  AAT GCC AAC CTA TGG AGA AAT TTC CTT TAC TCC AAT ATT GCG CTG TAC Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr 485  CTG CCA GAC AAG CTA AAA TAC AAC CCC ACC ACT AAT GTG GAA ATT TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 510  AAC CCC AAC ACC TAC GAC TAC ATC ATC AAC AAG CGA CTG GTG GCT CCC GGG Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 525  CTT GTA GAC TGC TAC ATT AAC CTT GGG GCG CGC TGG TC CTG GAC TAC Leu Val Asp Cyr Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CAC GAT GCG GCC CTC CGT Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 545  TAT CGC TCC ATG TTG TTG GGA AAC GGC CGC TAC GTG CCC TTT CAC ATT Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 560  TAT CGC TCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTC CTG CAA CIN Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro 580  GGC TCA TAT ACA TAT GAA TGG AAC TTC AGG AAG GAT GTT AAC ATG GTT CTC CTC CTC CTC CTC CTC CTC CTC C		GGC Gly	GAT Asp	Asn	GGA Gly	GAT Asp	ACT Thr	ACA Thr	Trp	ACA Thr	AAA Lys	GAT Asp	GAA Glu	Thr	TTT Phe	GCA Ala	ACA Thr	1392
Asn Ala Asn Leu Trp Arg Asn Phe Leu Tyr Ser Asn Ile Ala Leu Tyr 485  CTG CCA GAC AAG CTA AAA TAC AAC CCC ACC AAT GTG GAA ATA TCT GAC Leu Pro Asp Lys Leu Lys Tyr Asn Pro Thr Asn Val Glu Ile Ser Asp 500  AAC CCC AAC ACC TAC GAC TAC ATG ATG AAC AAG CGA GTG GTG GCT CCC GGG Asn Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 515  CTT GTA GAC TCC TAC ATT AAC CTT GGG GCG CGC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 540  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CGC AAT GCG GGC CTC CGT Asn His His Arg Asn Ala Gly Leu Arg 555  TAT CGC TCC ATG TTG TTG GGA AAC GGC CGC TAC GTG CCC TTT CAC ATT Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 560  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CAC TYR ASN GGG CGC CGC TAC GTG CCC TTT CAC ATT Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 575  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CCA 17  Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Pro 585  GGC TCA TAT ACA TAT GAA TGG AAC TTC AGG AAG GAT GTT AAC ATG GTT GIV Pro Sp5 S80  GGC TCA TAT ACA TAT GAA TGG AAC TCT AGA GTT GAC GGG GCT AGC ATT ASN Met Val Sp5 S85  CTG CAG AGC TCT CTG GGA AAC GAC CCC TTC TC CTC CTC CTC CTC CTC CTC CT		CGT Arg	Asn	GAA Glu	ATA Ile	GGA Gly	GTG Val	Gly	AAC Asn	AAC Asn	TTT Phe	GCC Ala	Met	GAA Glu	ATT Ile	AAC Asn	CTA Leu	1440
Leu Pro Asp Lys Leu Lys Tyr Asn Pro 505         Thr Asn Val Glu Ile Ser Asp 510           AAC CCC AAC ACC TAC GAC TAC ACT TAC ATG AAC AAG CGA GTG GTG GCT CCC GGG ASN Pro Asn Thr Tyr Asp Tyr Met Asn Lys Arg Val Val Ala Pro Gly 515         15           CTT GTA GAC TGC TAC ATT AAC CTT GGG GCG CGC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530         16           ATG GAC AAC GTT AAT CCC TTT AAC CAC CGC CGC AAT GCG GGC CTC CGT Met Asp Asn Val Asn Pro Phe Asn His His Arg Asn Ala Gly Leu Arg 5545         16           ATG CGC TCC ATG TTG TG GGA AAC GGC GCC TAC GTG TY Val Asp Ser Met Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile S75         17           TAT CGC TCC ATG TTG TTG GGA AAC GGC ATT TTT TTT GCC ATT TYR ARG Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 575         17           CAG GTG CCC CAA AAG TTT TTT GCC ATT TTT GCA ATT TYR GLY ASN Leu Leu Leu Leu Pro 580         17           GGT TCA TAT ACA TAT GAA TGG AAC TTC AGG AGG GAT GTT AAC ATG GTT GLY Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val 605         18           GGC TCA TAT ACA TAT GAA TGG AAC GAC CTT AGA GTT GAC GGG GCT AGC ATT Leu Gly Ser Tyr Thr Tyr Glu Trp Asn Asp Clu Arg Val Asp Gly Ala Ser Ile 600         18           CTG CAG AGC TCT CTG GGA AAC GAC CTT AGA GTT GAC GGG GCT AGC ATT Leu Arg Val Asp Gly Ala Ser Ile 600         18           CTG CAG AGC TCT CTG GGG AAC GAC CTT AGA GTT GAC GGG GCT AGC ATT Leu Arg Val Asp Gly Ala Ser Ile 600         18           CTG CAG AGC TCT CTG GGG AAC GAC CTC TCT CTG GAC ACC TTC TTC CCC ATG GCC CAC CAC CAC CAC CAC CAC CAC CAC CA		Asn	GCC Ala	AAC Asn	CTA Leu	TGG Trp	Arg	AAT Asn	TTC Phe	CTT Leu	TAC	Ser	AAT Asn	ATT Ile	GCG Ala	CTG Leu	Tyr	1488
Ash Pro Ash Thr Tyr Asp Tyr Met Ash Lys Arg Val Val Ala Pro Gly 515  THE TYR Asp Tyr Met Ash Leu 520  THE GAC GAC TGC TAC ATT AAC CTT GGG GGG GGC GGC TGG TCT CTG GAC TAC Leu Val Asp Cys Tyr Ile Ash Leu Gly Ala Arg Trp 540  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CGC AAT GGG GGC CTC CGT Met Asp Ash Val Ash Pro 550  ATG GAC AAC GTT AAT CCC TTT AAC CAC CAC CGC AAT GGG GGC CTC CGT Met Asp Ash Val Ash Pro 550  TAT CGC TCC ATG TTG TTG GGA AAC GGC CGC TAC Tyr Arg Ser Met Leu Leu Gly Ash Gly Arg Tyr Val Pro Phe His Ile 575  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CAC CGC TAC CGC TAC CTC CTC CTG CAC CGC TAC CGC TAC CTC CTC CTC CTC CTC CTC CTC CTC CT		CTG Leu	CCA Pro	GAC Asp	AAG Lys	Leu	AAA Lys	TAC Tyr	AAC Asn	CCC Pro	Thr	AAT Asn	GTG Val	GAA Glu	ATA Ile	Ser	GAC Asp	1536
Leu Val Asp Cys Tyr Ile Asn Leu Gly Ala Arg Trp Ser Leu Asp Tyr 530 Cs GAC AAC GTT AAT CCC TTT AAC CAC CAC CAC CGC AAT GCG GGC CTC CGT ASp Asn Val Asn Pro Ser Ser Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Ser Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Ser Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Ser Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Ser Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile Ser Arg CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CCA Ile CGln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Leu Leu Leu Leu Pro Ser Ser Leu Gly Asn Phe Arg Lys Asp Val Asn Met Val Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val Gly Ala Ser Ile Glo Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile Glo Ser Leu Gly Asn Cat CTC TTC CCC ATG CAC ATT Ile Cat CTC CTC CTC CTC CTC CTC CTC CTC CTC CT		AAC Asn	CCC Pro	AAC Asn	Thr	TAC Tyr	GAC Asp	TAC Tyr	ATG Met	Asn	AAG Lys	CGA Arg	GTG Val	GTG Val	Ala	CCC Pro	GGG Gly	1584
Met Asp Asn Val Asn Pro She Asn His His Arg Asn Ala Gly Leu Arg S45 Asn Val Asn Pro S50 Asn His His Arg Asn Ala Gly Leu Arg S55 Asn CGC TCC TCC CTG CCC TTT CAC ATT TYR Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 575  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CCA Leu Pro S80 Asn Leu Leu Leu Leu Pro S80 Asn Leu Leu Leu Leu Pro S80 Asn Leu Leu Leu Pro S90 Asn Met Val GOS Asn Met Val GOS Asn Asp CTT THR TYR Glu Trp Asn GOO Asn Asp CTT GAC GCT AGC ATT ASN ASP CON Met Ala His G25 ACC TCT CTC CTC CTG CCA ANG ATT ASN ASP CON Met Ala His G25 ACC ACC ASN ASP THR Asn Asp GCC ACC ASN ASP THR Asn Asp GCC CAC Asn ASP Thr Asn Asp G55 Asn Asp Thr Asn Asp G55 Asn Asp Thr Asn Asp G65 Asn Asp Thr Asn Asp Asp G65 Asn Asp Thr Asn A		CTT Leu	GTA Val	Asp	TGC Cys	TAC Tyr	ATT Ile	AAC Asn	Leu	GGG Gly	GCG Ala	CGC Arg	TGG Trp	Ser	CTG Leu	GAC Asp	TAC Tyr	1632
Tyr Arg Ser Met Leu Leu Gly Asn Gly Arg Tyr Val Pro Phe His Ile 575  CAG GTG CCC CAA AAG TTT TTT GCC ATT AAA AAC CTC CTC CTC CTG CCA 177  Gln Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Leu Pro 580  GGC TCA TAT ACA TAT GAA TGG AAC TTC AGG AAG GAT GTT AAC ATG GTT Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val 605  CTG CAG AGC TCT CTG GGA AAC GAT CTT AGA GTT GAC GGG GCT AGC ATT Leu Gln Ser Ser Leu Gly Asn Asp Asp Leu Arg Val Asp Gly Ala Ser Ile 610  AAG TTT GAC AGC ATT TGT CTT TAC GCC ACC TTC TTC CCC ATG GCC CAC Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 625  AAC ACG GCC TCC ACG CTG GAA GCC ATG CTC AGA AAT GAC ACC AAC GAC ASn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 655  CAG TCC TTT AAT GAC TAC CTT TCC GCC GCC AAC AAC ATG CTA TAC CCC ATA GIN Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		ATG Met	Asp	AAC Asn	GTT Val	AAT Asn	CCC Pro	Phe	AAC Asn	CAC His	CAC His	CGC Arg	Asn	GCG Ala	GGC Gly	CTC Leu	CGT Arg	1680
Gen Val Pro Gln Lys Phe Phe Ala Ile Lys Asn Leu Leu Leu Leu Leu Pro 580  Geo Tca Tat Aca Tat Gaa Teg Aac Ttc Aeg Aac Gat Gtt Aac Ate Gtt Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val 605  CTe Cae Aeg Tct Cte Gea Aac Gat ctt Aeg Gtt Gee Gct Aeg Ate Gtt Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile 610  Aac Ttt Gac Aeg Att Teg Ctt Tac Gcc Acc Ttc Ttc Ccc Ate Gcc Cac Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 625  Aac Acc Gcc Tcc Acc Cte Gaa Gcc Ate Ctc Aeg Aat Gac Aeg		Tyr	CGC Arg	TCC Ser	ATG Met	TTG Leu	Leu	Gly	AAC Asn	GGC Gly	CGC Arg	Tyr	GTG Val	CCC Pro	TTT Phe	CAC His	Ile	1728
Gly Ser Tyr Thr Tyr Glu Trp Asn Phe Arg Lys Asp Val Asn Met Val 595  CTG CAG AGC TCT CTG GGA AAC GAT CTT AGA GTT GAC GGG GCT AGC ATT Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile 610  AAG TTT GAC AGC ATT TGT CTT TAC GCC ACC TTC TTC CCC ATG GCC CAC Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 625  AAC ACG GCC TCC ACG CTG GAA GCC ATG CTC AGA AAT GAC ACC AAC GAC Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 640  CAG TCC TTT AAT GAC TAC CTT TCC GCC GCC AAC ATG CTA TAC CCC ATA Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		CAG Gln	GTG Val	CCC Pro	CAA Gln	Lys	TTT Phe	TTT Phe	GCC Ala	ATT Ile	Lys	AAC Asn	CTC Leu	CTC Leu	CTC Leu	Leu	CCA Pro	1776
Leu Gln Ser Ser Leu Gly Asn Asp Leu Arg Val Asp Gly Ala Ser Ile 610  AAG TTT GAC AGC ATT TGT CTT TAC GCC ACC TTC TTC CCC ATG GCC CAC Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 625  AAC ACG GCC TCC ACG CTG GAA GCC ATG CTC AGA AAT GAC ACC AAC GAC Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 640  CAG TCC TTT AAT GAC TAC CTT TCC GCC GCC AAC ATG CTA TAC CCC ATA Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		GGC Gly	TCA Ser	TAT Tyr	Thr	TAT Tyr	GAA Glu	TGG Trp	AAC Asn	Phe	AGG Arg	AAG Lys	GAT Asp	GTT Val	Asn	ATG Met	GTT Val	1824
Lys Phe Asp Ser Ile Cys Leu Tyr Ala Thr Phe Phe Pro Met Ala His 625  AAC ACG GCC TCC ACG CTG GAA GCC ATG CTC AGA AAT GAC ACC AAC GAC Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 640  CAG TCC TTT AAT GAC TAC CTT TCC GCC GCC AAC ATG CTA TAC CCC ATA Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		CTG Leu	CAG Gln	Ser	TCT Ser	CTG Leu	GGA Gly	AAC Asn	Asp	CTT Leu	AGA Arg	GTT Val	GAC Asp	Gly	GCT Ala	AGC Ser	ATT Ile	1872
Asn Thr Ala Ser Thr Leu Glu Ala Met Leu Arg Asn Asp Thr Asn Asp 640 655  CAG TCC TTT AAT GAC TAC CTT TCC GCC GCC AAC ATG CTA TAC CCC ATA Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		AAG Lys	Phe	GAC Asp	AGC Ser	ATT Ile	TGT Cys	Leu	TAC Tyr	GCC Ala	ACC Thr	TTC Phe	Phe	CCC Pro	ATG Met	GCC Ala	CAC His	1920
Gln Ser Phe Asn Asp Tyr Leu Ser Ala Ala Asn Met Leu Tyr Pro Ile		Asn	ACG Thr	GCC Ala	TCC Ser	ACG Thr	Leu	GAA Glu	GCC Ala	ATG Met	CTC Leu	Arg	AAT Asn	GAC Asp	ACC Thr	AAC Asn	Asp	1968
	1	CAG Gln	TCC Ser	TTT Phe	AAT Asn	Asp	TAC Tyr	CTT Leu	TCC Ser	GCC Ala	Ala	AAC Asn	ATG Met	CTA Leu	TAC Tyr	Pro	ATA Ile	2016

	٠.																
							Pro					TCG Ser					2064
												ACA Thr 700					2112
												TAC Tyr					2160
												ACC Thr					2208.
												GGC Gly					2256
									_			GTT Val					2304
												TGG Trp 780			GTG Val		2352
												TTC Phe				·	2400
												AAC Asn				,	2448
												GAG Glu					2496
		Ile										GTA Val					2544
												AAC Asn 860					2592
CCA Pro	CTA Leu 865	ATA Ile	GGC Gly	AAA Lys	ACC Thr	GCG Ala 870	GTT Val	GAC Asp	AGT Ser	ATT Ile	ACC Thr 875	CAG Gln	AAA Lys	AAG Lys	TTT Phe		2640
												AGT Ser					2688
												CTC Leu					.2736
												CCC Pro					2784

CCC	ACC	CTI Leu 930	тел	TAT Tyr	GTT Val	TTG Leu	TTT Phe 935	Glu	GTC Val	TTT Phe	GAC Asp	GTG Val 940	Val	CGT Arg	GTG Val		2832
CAC His	CAG Gln 945	CCG	CAC His	CGC Arg	GGC Gly	GTC Val 950	ATC Ile	GAG Glu	ACC Thr	GTG Val	TAC Tyr 955	CTG Leu	CGC Arg	ACG Thr	CCC		2880
TTC Phe 960	TCG Ser	GCC Ala	GGC	AAC Asn	GCC Ala 965	ACA Thr	ACA Thr	TAA									2,907
(2)	INF	ORMA	TION	FOR	SEQ	ID	NO: 5	2:									
		(	A) L B) T D) T	ENGT YPE: OPOL	H: 9 ami: OGY:	67 a no a lin	ear	CS: aci	ds								
	(ii)	MO	LECU	LE T	YPE:	pro	tein									•	
	(xi)	SE	QUEN	CE D	ESCR:	IPTI	: : NC	SEQ :	ID N	0:52	:						
1				5			Gln		10					15			
Gln	Asp	Ala	Ser 20	Glu	Tyr	Leu	Ser	Pro 25	Gly	Leu	Val	Gln	Phe 30	Ala	Arg		•
Ala	Thr	G1u 35	Thr	Tyr	Phe	Ser	Leu 40	Asn	Asn	Lys	Phe	Arg 45	Asn	Pro	Thr		
Val	Ala 50	Pro	Thr	His	Asp	Val 55	Thr	Thr	Asp	Arg	Ser 60	Gln	Arg	Leu	Thr		
Leu 65	Arg	Phe	Ile	Pro	Val 70	Asp	Arg	Glu	Asp	Thr 75	Ala	Tyr	Ser	Tyr	Lys 80	•	
Ala	Arg	Phe	Thr	Leu 85	Ala	Val	Gly	Asp	Asn 90	Arg	Val	Leu	Asp	Met 95	Ala	•	
Ser	Thr	Tyr	Phe 100	Asp	Ile	Arg	Gly	Val 105	Leu	Asp	Arg	Gly	Pro 110	Thr	Phe		
Lys	Pro	Tyr 115	Ser	Gly	Thr	Ala	Tyr 120	Asn	Ala	Leu	Ala	Pro 125	Lys	Gly	Ala		-
ro	Asn 130	Ser	Cys	Glu	Trp	Glu 135	Gln	Thr	Glu	Asp	Ser 140	Gly	Arg	Ala	Val		
11a .45	Glu	Asp	Glu	Glu	Glu 150	Glu	Asp	Glu	Asp	Glu 155	Glu	Glu	Glu	Glu	Glu 160		•
lu	Gln	Asn	Ala	Arg 165	Asp	Gln	Ala	Thr	Lys 170	Lys	Thr	His	Val	Tyr 175	Ala		
ln	Ala	Pro	Leu 180	Ser	Gly	Glu	Thr	Ile 185	Thr	Lys	Ser	Gly	Leu 190	Gln	Ile		

Gly Ser Asp Asn Ala Glu Thr Gln Ala Lys Pro Val Tyr Ala Asp Pro 195 200 205

Ser Tyr Gln Pro Glu Pro Gln Ile Gly Glu Ser Gln Trp Asn Glu Ala 210 215 220

Asp 225	Ala	Asn	Ala	Ala	Gly 230	Gly	Arg	Val	Leu	Lys 235	Lys	Thr	Thr	Pro	Met 240
Lys	Pro	Cys	Tyr	Glý 245	Ser	Tyr	Ala	Arg	Pro 250	Thr	Asn	Pro	Phe	Gly 255	Gly
Gln	Ser	Val	Leu 260	Val	Pro	Asp	Glu	Lys 265	Gly	Val	Pro	Leu	Pro 270	Lys ·	Val
Asp	Leu	Gln 275	Phe	Phe	Ser	Asn	Thr 280	Thr	Ser	Leu	Asn	Asp 285	Arg	Gln	Gly
Asn	Ala 290	Thr	Lys	Pro	Lys	Val 295	Val	Leu	Tyr	Ser	Glu 300	Asp	Val	Asn	Met
Glu 305	Thr	Pro	Asp	Thr	His 310	Leu	Ser	Tyr	Lys	Pro 315	·Gly	Lys	Gly	Asp	Glu 320
Asn	Ser	Lys	Ala	Met 325	Leu	Gly	Gln	Gln	Ser 330	Met	Pro	Asn	Arg	Pro 335	Asn
Tyr	Ile	Ala	Phe 340	Arg	Asp	Asn	Phe	Ile 345	Gly	Leu	Met	Tyr	Tyr 350	Asn	Ser
Thr	Gly	Asn 355	Met	Gly	Val	Leu	Ala 360	Gly	Gln	Ala	Ser	Gln 365	Leu	Asn	Ala
Val	Val 370	Asp	Leu	Gln	Asp	Arg 375	Asn	Thr	Glu	Leu	Ser 380	Tyr	Gĺn	Leu	Leu.
Leu 385	Asp	Ser	Ile	Gly	Asp 390	Arg	Thr	Arg	Tyr	Phe 395	Ser	Met	Trp	Asn	Gln 400
Ala	Val	Asp	Ser	Tyr 405	Asp	Pro	Asp	Val	Arg 410	Ile	Ile	Glu	Asn	His 415	Gly
Thr	Glu	Asp	Glu 420	Leu	Pro	Asn	Tyr	Cys 425	Phe	Pro	Leu	Gly	Gly 430	Ile	Gly
Val	Tḥr	Asp 435	Thr	Tyr	Gln	Ala	Ile 440	Lys	Ala	Asn	Gly	Asn 445	Gly	Ser	Gly
Asp	Asn 450	Gly	Asp	Thr	Thr	Trp 455	Thr	Lys	Asp	Glu	Thr 460	Phe	Ala	Thr	Arg
Asn 465	Glu	Ile	Gly	Val	Gly 470	Asn	Asn	Phe	Ala	Met 475	Glu	Ile	Asn	Leu	Asn 480
Ala	Asn	Leu	Trp	Arg 485	Asn	Phe	Leu	Tyr	Ser 490	Asn	Ile	Ala	Leu	Tyr 495	Leu
Pro	Asp	Lys	Leu 500	Lys	Tyr	Asn	Pro	Thr 505	Asn	Val	Glu	Ile	Ser 510	Asp	Asn
Pro	Asn	Thr 515	Tyr	Asp	Tyr	Met	Asn 520	Lys	Arg	Val	Val	Ala 525	Pro	Gly	Leu
Val	Asp 530	Суѕ	Tyr	Ile	Asn	Leu 535	Gly	Ala	Arg	Trp	Ser 540	Leú	Asp	Tyr	Met
Asp 545		Val	Asn	Pro	Phe 550	Asn	His	His	Arg	Asn 555	Ala	Gly	Leu	Arg	Tyr .

Arg	Ser	. Met	Leu	Leu 565	Gly	Asn	Gly	Arg	Tyr 570	Val	Pro	Phe	His	575	
Val	Pro	Gln	Lys 580	Phe	Phe	Ala	Ile	Lys 585	Asn	Leu	Leu	Lev	Lev 590		Gl
Ser	Tyr	Thr 595	Tyr	Glu	Trp	Asn	Phe 600	Arg	Lys	Asp	Val	Asn 605		Val	Lei
Gln	Ser 610	Ser	Leu	Gly	Asn	Asp 615	Leu	Arg	Val	Asp	Gly 620		Ser	Ile	Lys
023		•			Leu 630					635					640
				645	Glu				650					655	
			000		Leu			665	•				670		
		6/5			Val		680					685			
	030				Ala	695					700				
/03					Tyr 710					715					720
				125	Thr				730					735	
			740		Ser			745					750		
		/55			Phe		760					765			_
	770				Cys	115					780				
03					Asn 790					795					800
				805	Met				810					815	
			820		Asp			825					830		
		033			His .		840					845			
	030					855					860				
05					Ala 870					875					880
ys	Asp	Arg	Thr	Leu 885	Trp .	Arg	Ile	Pro	Phe 890	Ser	Ser	Asn	Phe	Met 895	Ser

									-							
Met	Gly	Ala	Leu 900	Thr	Asp	Leu	Gly	Gln 905	Asn	Leu	Leu	Tyr	Ala 910	Asn	Ser	. •
Ala	His	Ala 915	Leu	Asp	Met	Thr	Phe 920	Glu	Val	Asp	Pro	Met 925	Asp	Glu	Pro	•
Thr	Leu 930	Leu	Tyr	Val	Leu	Phe 935	Glu	Val	Phe	Asp	Val 940	Val	Arg	Val	His	
Gln 945	Pro	His	Arg	Gly	Val 950	Ile	Glu	Thr	Val	Tyr 955	Leu	Arg	Thr	Pro	Phe 960	
Ser	Ala	Gly	Asn ·	Ala 965	Thr	Thr			:							
(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:53	3: -								
	(i)	( <i>F</i> (E	A) LE 3) TY C) SI	NGTH PE: RANI	I: 28 nucl	CTERI 358 k leic ESS: line	ase acid	pai:	cs							
	(ii)	MOI	ECUI	E TY	PE:	DNA	(ger	nomic	<b>)</b>							•
	(xi)	SEC	QUENC	CE DE	SCRI	PTIC	on: s	SEQ 1	D NO	53:						
ATG													CAC His			48
GGC Gly	CAG Gln	GAC Asp	GCC Ala	TCG Ser 20	GAG Glu	TAC Tyr	CTG Leu	AGC Ser	CCC Pro 25	GGG Gly	CTG Leu	GTG Val	CAG Gln	TTT Phe 30	GCC Ala	96
													AGA Arg 45			144
													CAG Gln			192
		Arg	Phe	Ile	Pro	Val	Asp	Arg		Asp		Ala	TAC Tyr			240
													CTG Leu			288
GCT Ala	TCC Ser	ACG Thr	TAC Tyr	TTT Phe 100	GAC Asp	ATC Ile	CGC Arg	GGC Gly	GTG Val 105	CTG Leu	GAC Asp	AGG Arg	GGC Gly	CCT Pro 110	ACT Thr	336
													CCC Pro 125			384
													CTT Leu			432
AAC	CTA	GAA	GAA	GAG	GAC	GAT	GAC	AAC	GAA	GAC	GAA	GTA	GAC	GAG	CAA	480

Asn	Leu 145	Glu	Glu	Glu	Asp	Asp 150	Asp	Asn	Glu	Asp	Glu 155		Asp	Glu	Gln	
GCT Ala 160	GIU	CAG Gln	CAA Gln	AAA Lys	ACT Thr 165	His	GTA Val	TTT Phe	GGG	CAG Gln 170	Ala	CCT Pro	TAT Tyr	TCT Ser	GGT Gly 175	528
ATA	AAT Asn	ATT Ile	ACA Thr	AAG Lys 180	GLu	GGT Gly	ATT Ile	CAA Gln	ATA Ile 185	GGT Gly	GTC Val	GAA Glu	GGT Gly	CAA Gln 190	ACA Thr	576
CCT Pro	AAA Lys	TAT Tyr	GCC Ala 195	Asp	AAA Lys	ACA Thr	TTT Phe	CAA Gln 200	Pro	GAA Glu	CCT Pro	CAA Gln	ATA Ile 205	GGA Gly	GAA Glu	624
TCT Ser	CAG Gln	TGG Trp 210	TAC Tyr	GAA Glu	ACT Thr	GAA Glu	ATT Ile 215	AAT Asn	CAT His	GCA Ala	GCT Ala	GGG Gly 220	AGA Arg	GTC Val	CTT Leu	672
гÃ2	225	THE	Thr	.Pro	Met	Lys 230	Pro	Cys	TAC Tyr	Gly	Ser 235	Tyr	Ala	Lys	Pro	720
240	ASII	GIU	ASI	стА	245	GIn	Gly	Ile	CTT Leu	Val 250	Lys	Gln	Gln	Asn	Gly 255	768
AAG Lys	CTA Leu	GAA Glu	AGT Ser	CAA Gln 260	GTG Val	GAA Glu	ATG Met	CAA Gln	TTT Phe 265	TTC Phe	TCA Ser	ACT Thr	ACT Thr	GAG Glu 270	GCG Ala	816
ACC Thr	GCA Ala	GGC Gly	AAT Asn 275	GGT Gly	GAT Asp	AAC Asn	TTG Leu	ACT Thr 280	CCT Pro	AAA Lys	GTG Val	GTA Val	TTG Leu 285	TAC Tyr	AGT Ser	864
GAA Glu	GAT Asp	GTA Val 290	GAT Asp	ATA Ile	GAA Glu	ACC Thr	CCA Pro 295	GAC Asp	ACT Thr	CAT His	ATT Ile	TCT Ser 300	TAC Tyr	ATG Met	CCC Pro	912
ACT Thr	ATT Ile 305	AAG Lys	GAA Glu	GGT Gly	AAC Asn	TCA Ser 310	CGA Arg	GAA Glu	CTA Leu	ATG Met	GGC Gly 315	CAA Gln	CAA Gln	TCT Ser	ATG Met	960
CCC Pro 320	AAC Asn	AGG Arg	CCT Pro	AAT Asn	TAC Tyr 325	ATT Ile	GCT Ala	TTT Phe	AGG Arg	GAC Asp 330	AAT Asn	TTT Phe	ATT Ile	GGT Gly	CTA Leu 335	1008
ATG Met	TAT Tyr	TAC Tyr	AAC Asn	AGC Ser 340	ACG Thr	GGT Gly	AAT Asn	ATG Met	GGT Gly 345	GTT Val	CTG Leu	GCG Ala	GGC Gly	CAA Gln 350	GCA Ala	1056
rcg Ser	CAG Gln	TTG Leu	AAT Asn 355	GCT Ala	GTT Val	GTA Val	GAT Asp	TTG Leu 360	CAA Gln	GAC Asp	AGA Arg	AAC Asn	ACA Thr 365	GAG Glu	CTT Leu	1104
rca Ser	IYE	CAG Gln 370	CTT Leu	TTG Leu	CTT Leu	Asp	TCC Ser 375	ATT Ile	GGT Gly	GAT Asp	AGA Arg	ACC Thr 380	AGG Arg	TAC Tyr	TTT Phe	1152
oet	ATG Met 385	TGG Trp	AAT Asn	CAG Gln	GCT Ala	GTT Val 390	GAC Asp	AGC Ser	TAT Tyr	GAT Asp	CCA Pro 395	GAT Asp	GTT Val	AGA Arg	ATT Ile	1200
TTA	GAA .	AAT	CAT	GGA .	ACT	GAA	GAT	GAA	CTT	CCA	AAT	TAC	TGC	TTT	CCA	1248

Ile 400	Glu	Asn	His	Gly	Thr 405	Glu	Asp	Glu	Leu	Pro 410	Asn	Tyr	Суз	Phe	Pro 415		
						ACA Thr											1296
						TGG Trp											1344
						GGA Gly										,	1392
						AAT Asn 470											1440
						TAC Tyr											1488
						TAC Tyr										•	1536
						AAC Asn										•	1584
						TTT Phe							•				1632
						GGC Gly 550											1680
						TTT Phe										,	1728
						TGG Trp											1776
						AAT Asn											1824
						CTT Leu											1872
						GAG Glu 630											1920
						CTC Leu											1968

									50							
CCC Pro	GCC Ala	AAC Asn	GCT Ala	ACC Thr 660	AAC Asn	GTG Val	CCC Pro	ATA Ile	Ser 665	ATC Ile	CCC Pro	TCC Ser	CGC Arg	AAC Asn 670	TGG	2016
GCG Ala	GCT Ala	TTC Phe	CGC Arg 675	GGC Gly	TGG Trp	GCC Ala	TTC Phe	ACG Thr 680	CGC Arg	CTT Leu	AAG Lys	ACT Thr	AAG Lys 685	GAA Glu	ACC Thr	2064
CCA Pro	TCA Ser	CTG Leu 690	Gly	TCG Ser	GGC Gly	TAC Tyr	GAC Asp 695	CCT Pro	TAT Tyr	TAC Tyr	ACC Thr	TAC Tyr 700	TCT Ser	GGC Gly	TCT Ser	2112
ATA Ile	CCC Pro 705	TAC Tyr	CTA Leu	GAT Asp	GGA Gly	ACC Thr 710	TTT Phe	TAC Tyr	CTC Leu	AAC Asn	CAC His 715	ACC Thr	TTT Phe	AAG Lys	AAG Lys	2160
GTG Val 720	GCC Ala	ATT Ile	ACC Thr	TTT Phe	GAC Asp 725	TCT Ser	TCT Ser	GTC Val	AGC Ser	TGG Trp 730	CCT	GGC Gly	AAT Asn	GAC Asp	CGC Arg 735	2208
CTG Leu	CTT Leu	ACC Thr	CCC Pro	AAC Asn 740	GAG Glu	TTT Phe	GAA Glu	ATT Ile	AAG Lys 745	CGC Arg	TCA Ser	GTT Val	GAC Asp	GGG Gly 750	GAG Glu	2256
GGT Gly	TAC Tyr	AAC Asn	GTT Val 755	GCC Ala	CAG Gln	TGT Cys	AAC Asn	ATG Met 760	ACC Thr	AAA Lys	GAC Asp	TGG Trp	TTC Phe 765	CTG Leu	GTA Val	2304
CAA Gln	ATG Met	CTA Leu 770	GCT Ala	AAC Asn	TAC Tyr	AAC Asn	ATT Ile 775	GGC Gly	TAC Tyr	CAG Gln	GGC Gly	TTC Phe 780	TAT Tyr	ATC Ile	CCA Pro	2352
GAG Glu	AGC Ser 785	TAC Tyr	AAG Lys	GAC Asp	CGC Arg	ATG Met 790	TAC Tyr	TCC Ser	TTC Phe	TTT Phe	AGA Arg 795	AAC Asn	TTC Phe	CAG Gln	CCC Pro	2400
ATG Met 800	AGC Ser	CGT Arg	CAG Gln	GTG Val	GTG Val 805	GAT Asp	GAT Asp	ACT Thr	AAA Lys	TAC Tyr 810	AAG Lys	GAC Asp	TAC Tyr	CAA Gln	CAG Gln 815	2448
GTG Val	GGC Gly	ATC Ile	CTA Leu	CAC His 820	CAA Gln	CAC His	AAC Asn	AAC Asn	TCT Ser 825	GGA Gly	TTT Phe	GTT Val	GGC Gly	TAC Tyr 830	CTT Leu	2496
GCC Ala	CCC Pro	ACC Thr	ATG Met 835	CGC Arg	GAA Glu	GGA Gly	CAG Gln	GCC Ala 840	TAC Tyr	CCT Pro	GCT Ala	AAC Asn	TTC Phe 845	CCC Pro	TAT Tyr	2544
CCG Pro	CTT Leu	ATA Ile 850	GGC Gly	AAG Lys	ACC Thr	GCA Ala	GTT Val 855	GAC Asp	AGC Ser	ATT Ile	ACC Thr	CAG Gln 860	AAA Lys	AAG Lys	TTT Phe	2592
CTT Leu	TGC Cys 865	GAT Asp	CGC Arg	ACC Thr	CTT Leu	TGG Trp 870	CGC Arg	ATC Ile	CCA Pro	TTC Phe	TCC Ser 875	AGT Ser	AAC Asn	TTT Phe	ATG Met	2640
TCC Ser 880	ATG Met	GGC Gly	GCA Ala	Leu	ACA Thr 885	GAC Asp	CTG Leu	GGC Gly	CAA Gln	AAC Asn 890	CTT Leu	CTC Leu	TAC Tyr	GCC Ala	AAC Asn 895	2688
TCC Ser	GCC Ala	CAC His	GCG Ala	CTA Leu 900	GAC Asp	ATG Met	ACT Thr	TTT Phe	GAG Glu 905	GTG Val	GAT Asp	CCC Pro	ATG Met	GAC Asp 910	GAG Glu	2736

 		 TAT Tyr							_		2784
 	-	CGC Arg		_			 				2832
 		 AAC Asn	 		AA			·			2858

- (2) INFORMATION FOR SEQ ID NO:54:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 951 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

Ala Thr Pro Ser Met Met Pro Gln Trp Ser Tyr Met His Ile Ser Gly

Gln Asp Ala Ser Glu Tyr Leu Ser Pro Gly Leu Val Gln Phe Ala Arg

Ala Thr Glu Thr Tyr Phe Ser Leu Asn Asn Lys Phe Arg Asn Pro Thr

Val Ala Pro Thr His Asp Val Thr Thr Asp Arg Ser Gln Arg Leu Thr

Leu Arg Phe Ile Pro Val Asp Arg Glu Asp Thr Ala Tyr Ser Tyr Lys

Ala Arg Phe Thr Leu Ala Val Gly Asp Asn Arg Val Leu Asp Met Ala

Ser Thr Tyr Phe Asp Ile Arg Gly Val Leu Asp Arg Gly Pro Thr Phe 100 105

Lys Pro Tyr Ser Gly Thr Ala Tyr Asn Ala Leu Ala Pro Lys Gly Ala

Pro Asn Pro Cys Glu Trp Asp Glu Ala Ala Thr Ala Leu Glu Ile Asn 135 140

Leu Glu Glu Asp Asp Asp Asn Glu Asp Glu Val Asp Glu Gln Ala

Glu Gln Gln Lys Thr His Val Phe Gly Gln Ala Pro Tyr Ser Gly Ile

Asn Ile Thr Lys Glu Gly Ile Gln Ile Gly Val Glu Gly Gln Thr Pro 180

Lys Tyr Ala Asp Lys Thr Phe Gln Pro Glu Pro Gln Ile Gly Glu Ser 200

Gln Trp Tyr Glu Thr Glu Ile Asn His Ala Ala Gly Arg Val Leu Lys

Lys 225	Thi	Thi	r Pro	Met	230	Pro	Суз	Туг	Gl	235	Туг	Ala	Lys	Pro	240
Asr	ı Glu	a Asr	ı Gly	Gly 245	/ Gln	Gly	lle	Leu	Val 250	Lys	Gln	Glr	a Asn	Gly 255	
Leu	Glu	Ser	Glr. 260	Val	. Glu	Met	Gln	Phe 265	Phe	Ser	Thr	Thr	Glu 270		Thr
Ala	Gly	Asr 275	Gly	Asp	Asn	Leu	Thr 280	Pro	Lys	Val	Val	Leu 285		Ser	Glu
Asp	Val 290	Asp	Ile	Glu	Thr	Pro 295	Asp	Thr	His	Ile	Ser 300		Met	Pro	Thr
303					Ser 310					315				•	320
				325					330					335	
			340		Gly			345					350		
		333			Val		360					365		•	
	370					.375					380				,
363					Val 390					395					400
				405	Glu				410					415	
			420		Thr			425					430		
		433			Trp		440					445			
	450				Gly	455					460				
4,03					Asn 470					475					480
				485	Tyr				490					495	•
			500		Tyr			505					510		
		212			Asn		520					525			
	330				Phe	535					540				
Arg 545	Ser	Met	Leu	Leu	Gly 550	Asn	Gly	Arg	Tyr	Val 555	Pro	Phe	His	Ile	Gln 560

Val	Pro	Gln	Lys	Phe 565	Phe	Ala	Ile	Lys	Asn 570	Leu	Leu	Leu	Leu	Pro <b>5</b> 75	Gly
Ser	Tyr	Thr	Tyr 580	Glu	Trp	Asn	Phe	Arg 585	Lys	Asp	Val	Asn	Met 590	Val	Leu
Gln	Ser	Ser 595	Leu	Gly	Asn	Asp	Leu 600	Arg	Val	Asp	Gly	Ala 605	Ser	Ile	Lys
Phe	Asp 610	Ser	Ile	Cys	Leu	Tyr 615	Ala	Thr	Phe	Phe	Pro 620	Met	Ala	His	Asn
Thr 625	Ala	Ser	Thr	Leu	Glu 630	Ala	Met	Leu	Arg	Asn 635	Asp	Thr	Asn	Asp	Gln 640
Ser	Phe	Asn	Asp	Tyr 645	Leu	Ser	Ala	Ala	Asn 650	Met	Leu	Tyr	Pro	Ile. 655	Pro
Ala	Asn	Ala	Thr 660	Asn	Val	Pro		Ser 665	Ile	Pro	Ser	Arg	Asn 670	Trp	Ala
Ala ·	Phe	Arg 675	Gly	Trp	Ala	Phe	Thr 680	Arg	Leu	Lys	Thr	Lys 685	Glü	Thr	Pro
Ser	Leu 690	Gly	Ser	Gly	Tyr	Asp 695	Pro	Tyr	Tyr	Thr	Tyr- 700	Ser	Gly	Ser	Île
Pro 705	Tyr	Leu	Asp	Gly	Thr 710	Phe	Tyr	Leu	Asn	His 715	Thr	Phe	Lys	Lys	Val 720
Ala	Ile	Thr	Phe	Asp 725	Ser	Ser	Val		Trp 730	Pro	Gly	Asņ	Asp	Arg 735	Leu
Leu	Thr	Pro	Asn 740	Glu	Phe	Glu	Ile	Lys 745	Arg	Ser	Val	Asp	Gly 750	Glu	Gly
Tyr	Asn	Val 755	Ala	Gln	Cys	Asn	Met 760	Thr	Lys	Asp	Trp	Phe 765	Leu	Val	Gln
Met	Leu 770 <sub>.</sub>	Ala	Asn	Tyr	Asn	Ile 775	Gly	Tyr	Gln	Gly	Phe 780	Tyr	Ile	Pro	Glu
Ser 785	Tyr	Lys	Asp	Arg	Met 790	Tyr	Ser	Phe	Phe	Arg 795	Asn	Phe	Gln	Pro	Met 800
Ser	Arg	Gln	Val	Val 805	Asp	Asp	Thr	Lys	Tyr 810	Lys	Asp	Tyr	Gln	Gln 815	Val
Gly	Ile	Leu	His 820	Gln	His	Asn	Asn	Ser 825	Gly	Phe	Val	Gly	Tyr 830	Leu	Ala
Pro	Thr	Met 835	Arg	Glu	Gly	Gln	Ala 840	Tyŗ	Pro	Ala	Asn	Phe 845	Pro	Tyr	Pro
Leu	11e 850	Gly	Lys	Thr	Ala	Val 855	Asp	Ser	Ile	Thr	Gln 860	Lys	Lys	Phe	Leu
Cys 865	Asp	Arg	Thr	Leu	Trp 870	Arg	Ile	Pro	Phe	Ser 875	Ser	Asn	Phe	Met-	Ser 880
Met	Gly	Ala	Leu	Thr 885	Asp	Leu	Gly	Gln	Asn 890	Leu	Leu	Tyr	Ala	Asn 895	Ser

WO 98/40509

#### PCT/US98/05033

Ala	His	Ala	Leu	Asp	Met	Thr	Phe	Glu	Val	Asp	Pro	Met	Asp	Glu	Pro
			900					905		-			91 <b>0</b>		

Thr Leu Leu Tyr Val Leu Phe Glu Val Phe Asp Val Val Arg Val His 915 920 925

Arg Pro His Arg Gly Val Ile Glu Thr Val Tyr Leu Arg Thr Pro Phe 930 935 940

Ser Ala Gly Asn Ala Gln His 945 950

- (2) INFORMATION FOR SEQ ID NO:55:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 98 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
    - (ii) MOLECULE TYPE: other nucleic acid
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

GAA CTC GGA GGT GGA GGT GGA ACT AGT TTT GGA CGC GGA GAC ATT CGC
Glu Leu Gly Gly Gly Gly Thr Ser Phe Gly Arg Gly Asp Ile Arg
1 5 10 15

AAT TAAAGTACTG GATTCATGAC TCTAGACTTA ATTAAGGATC CAATAAA 98

(2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 17 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

Glu Leu Gly Gly Gly Gly Thr Ser Phe Gly Arg Gly Asp Ile Arg
1 10 15

Asn

WO 98/40509 PCT/US98/05033

103

#### WHAT IS CLAIMED IS:

1. A chimeric adenovirus coat protein comprising a nonnative amino acid sequence, wherein said chimeric adenovirus coat protein has a decreased ability or inability to be recognized by a neutralizing antibody directed against the wild-type adenovirus coat protein.

- 2. The chimeric adenovirus coat protein of claim 1, wherein said nonnative amino acid sequence comprises a deletion, insertion, or a replacement of a region of from about 1 to about 750 amino acids of said wild-type adenovirus coat protein.
- 3. The chimeric adenovirus coat protein of claim 1 or 2, wherein said nonnative amino acid sequence comprises a plurality of deletions, insertions, and/or replacements.
- 4. The chimeric adenovirus coat protein of any of claims 1-3, wherein said coat protein is a chimeric adenovirus hexon protein.
- 5. The chimeric adenovirus coat protein of claim 4, wherein said region deleted or replaced comprises a hypervariable region in either the 11 loop or the 12 loop.
- 6. The chimeric adenovirus coat protein of claim 5, wherein said hypervariable region is selected from the group consisting of HVR1, HVR2, HVR3, HVR4, HVR5, HVR6, and HVR7.
- 7. The chimeric adenovirus coat protein of any of claims 1-6, comprising a sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID

- NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44, SEQ ID NO:46, and SEQ ID NO:48.
- 8. The chimeric adenovirus coat protein of any of claims 1-7, wherein said nonnative amino acid sequence comprises a spacer of about 1 to about 750 amino acids.
- 9. The chimeric coat adenovirus coat protein of claim 8, wherein said spacer comprises the sequence of SEQ ID NO:50.
- 10. The chimeric adenovirus coat protein of any of claims 1-9, comprising an amino acid sequence of a coat protein of another serotype of adenovirus.
- 11. The chimeric adenovirus coat protein of claim 10, wherein said coat protein of another serotype is a hexon protein.
- 12. An isolated or purified nucleic acid that encodes the chimeric adenovirus coat protein of any of claims 1-11.
- 13. The isolated or purified nucleic acid of claim 12 comprising a sequence selected from the group consisting of SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:37, SEQ ID NO:39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:45, and SEQ ID NO:47.

WO 98/40509 PCT/US98/05033

- 14. The isolated or purified nucleic acid of claim 12 or 13 comprising SEQ ID NO:49.
- 15. An adenoviral vector that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 16. A method of genetically modifying a cell which comprises contacting said cell with the adenoviral vector of claim 15.
- 17. A host cell that comprises the chimeric adenovirus coat protein of any of claims 1-11.
- 18. A method of constructing an adenoviral vector that has a decreased ability or inability to be recognized by a neutralizing antibody directed against wild-type adenovirus coat protein, which method comprises obtaining an adenoviral vector comprising a wild-type adenovirus coat protein and replacing said wild-type adenovirus coat protein with the chimeric adenovirus coat protein of any of claims 1-11.

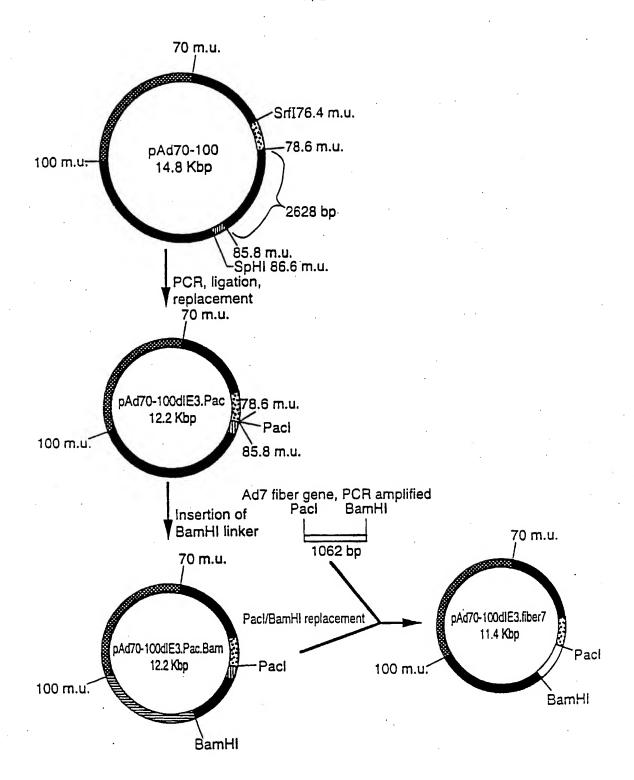


FIG. 1

SUBSTITUTE SHEET (RULE 26)

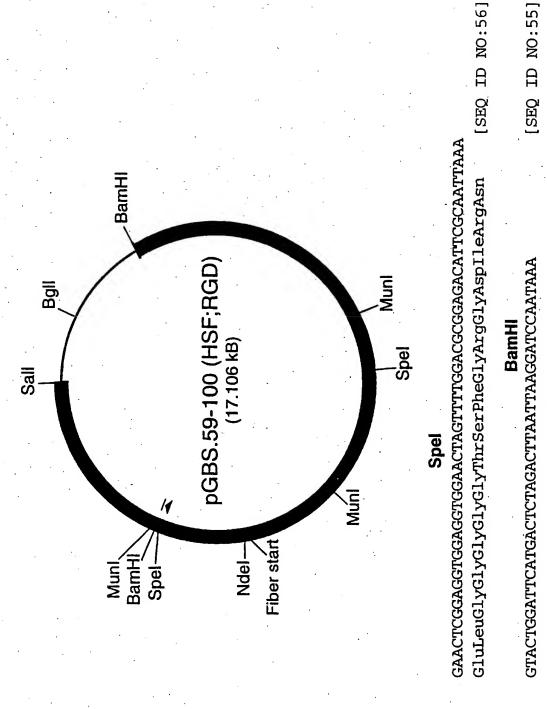


FIG. 2

# INTERNATIONAL SEARCH REPORT

Int .tional Application No PCT/US 98/05033

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IPC 6	IFICATION OF SUBJECT MATTER C12N15/86 C07K14/075 C12N15/	'34 C12N5/10						
According t	o International Patent Classification(IPC) or to both national classific	eation and IPC						
	SEARCHED							
Minimum d	ocumentation searched (classification system followed by classificat	alodmys nois						
IPC 6	C12N C07K							
Documenta	tion searched other than minimum documentation to the extent that	such documents are included in the fields searched						
Electronic o	lata base consulted during the international search (name of data b	ase and, where practical, search terms used)						
•								
	ENTS CONSIDERED TO BE RELEVANT							
Category °	Citation of document, with indication, where appropriate, of the re	levant passages Relevant to claim No.						
A ·	WO 96 26281 A (GENVEC, INC.) 29 1996	August 1-18						
	see page 5, line 7 - line 23 see page 6, line 30 - line 37 							
Α	CROMPTON J ET AL.: "Expression forein epitope on the surface of adenovirus hexon" JOURNAL OF GENERAL VIROLOGY, vol. 75, no. 1, January 1994, RE, pages 133-139, XP002071015 cited in the application see table 1	the						
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	er documents are listed in the continuation of box C.	X Patent family members are listed in annex.						
	egories of cited documents :	"T" later document published after the international filling date						
conside	nt defining the general state of the art which is not ared to be of particular relevance	or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention						
"L" documer	eanier document but published on or after the international filing date  "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to							
which is clied to establish the publication date of another citation or other special reason (as specified)  "Y" document of particular relevance; the claimed invention								
omer m		document is combined with one or more other such docu- ments, such combination being obvious to a person skilled in the art.	j					
later th	nt published prior to the international filing date but an the priority date claimed	"&" document member of the same patent family						
Date of the a	ctual completion of theinternational search	Date of mailing of the international search report						
9	July 1998	27/07/1998						
Name and m	alling address of the ISA  European Patent Office, P.B. 5818 Patentiaan 2	Authorized officer	$\neg$					
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in. utional Application No PCT/US 98/05033

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Category *	Citation of document, with indication where appropriate, of the relevant passages	Relevant to claim No.		
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